

IKKs and tumor cell plasticity

Serkan I. Göktuna^{1,2} , Michaela A. Diamanti³ and Tieu Lan Chau¹¹ Department of Molecular Biology and Genetics, Bilkent University, Ankara, Turkey² National Nanotechnology Research Center (UNAM), Bilkent University, Ankara, Turkey³ Georg-Speyer-Haus, Institute for Tumor Biology and Experimental Therapy, Frankfurt am Main, Germany

Keywords

EMT; I κ B kinases; IKK; inflammation; metastasis; Stemness; tumor cell plasticity

Correspondence

S. I. Göktuna, Department of Molecular Biology and Genetics, Bilkent University, 06800 Bilkent, Ankara, Turkey
Fax: +90 3122665097
Tel: +90 3122902418
E-mail: serkan.goktuna@bilkent.edu.tr

(Received 28 November 2017, revised 22 February 2018, accepted 21 March 2018)

doi:10.1111/febs.14444

Nuclear factor κ B (NF- κ B) transcription factors are the central hubs of signaling pathways connecting proinflammatory signals to cell survival, proliferation and cytokine production. In cancers, NF- κ B signaling influences many aspects of tumor development, from initiation to metastasis. These functions are mediated by tumor-induced plasticity that allows tumor cells to adapt and survive in changing conditions within the tumor microenvironment. Tumor cell plasticity is shaped by the inflammatory microenvironment in tumors. This review focuses on inhibitor of NF- κ B kinases, the direct upstream elements of NF- κ B regulation, specifically on their conventional and non-conventional functions in animal models of tumorigenesis from the recent literature.

Introduction

A relationship between inflammation and cancer has been proposed since Rudolph Virchow observed an increase in the number of infiltrating leukocytes in tumors in 1863 [1]. Since then, a large body of evidence has accumulated supporting such a link, and the underlying molecular mechanisms have gradually been uncovered. Today we know that a history of chronic inflammation can be attributed to more than 17% of malignancies worldwide [2]. Prolonged intake of non-

steroidal anti-inflammatory drugs significantly lowers the risk of certain cancers, such as breast and colon cancer [3], suggesting that therapies targeting inflammatory processes rather than directly killing tumor cells may overcome complications due to resistance of tumors to current therapeutics.

Drug resistance is acquired from cellular heterogeneity within tumors, which in turn arises from tumor cell plasticity. The mechanisms leading to tumor cell

Abbreviations

ATM1, ataxia telangiectasia mutated serine/threonine kinase 1; BAFF, B-cell activating factor; CAC, colitis-associated colorectal cancer; CAF, cancer-associated fibroblast; CaP, prostate cancer; CRC, colorectal cancer; CYLD, cylindromatosis lysine 63 deubiquitinase; DEN, diethylnitrosamine; DMBA, 7,12-dimethylbenzanthracene; E2F1, E2F transcription factor 1; EGFR, epithelial growth factor receptor; EMT, epithelial–mesenchymal transition; ER, endoplasmic reticulum; ERK, extracellular signal-regulated kinase; FOXA2, forkhead box protein A2; FOXO3a, forkhead box protein O3a; HCC, hepatocellular carcinoma; IFN, interferon; IKK, inhibitor of κ B kinase; IL, interleukin; I κ B, inhibitor of nuclear factor κ B; JNK, c-Jun N-terminal kinase; LT, lymphotoxin; MAPK, mitogen-activated protein kinase; mTOR, mechanistic target of rapamycin; NEMO, NF- κ B essential modulator; NF- κ B, nuclear factor κ B; NMSC, non-melanoma skin cancer; NPM, nucleophosmin; Rac1, Rac Family Small GTPase 1; RANK, receptor activator of nuclear factor κ B; RIPK1, receptor-interacting serine/threonine-protein kinase 1; ROS, reactive oxygen species; SCC, squamous cell carcinoma; SMRT, silencing mediator of retinoic acid and thyroid hormone receptor; SOD, superoxide dismutase; STAT3, signal transducer and activator of transcription 3; TAX1BP1, Tax1 binding protein 1; TGF β , tumor growth factor beta; TNF, tumor necrosis factor; TNFR, tumor necrosis factor receptor; TPA, 12-*O*-tetradecanoylphorbol-13-acetate; TSC, tuberous sclerosis complex subunit; VEGF-A, vascular endothelial growth factor A; XBP1, X-box-binding protein 1.

plasticity include mutations, epithelial–mesenchymal transition (EMT), dedifferentiation and inflammation, which help cancer cells adapt to changes and threats within the tumor microenvironment. Among these mechanisms, inflammation plays a central role. As will be explained in detail, the inflammatory microenvironment provides all the conditions for cells to accumulate further mutations, to dedifferentiate to gain stem cell-like properties and to go through the EMT for invasion and formation of distant metastases. Therefore, we need a better understanding of inflammatory pathways and their regulators to be able to develop superior therapeutic options that target tumor cell plasticity and tumor development. This review particularly focuses on inhibitor of nuclear factor κ B (I κ B) kinases (IKKs) as the master regulators of inflammatory signaling mechanisms that shape tumor cell plasticity in various cancer models.

Tumor cell plasticity

Plasticity by definition means the ability to change or adapt to varying conditions. Cellular plasticity is required for tumor cells to adapt to an ever-changing tumor microenvironment for growth and survival amidst various environmental threats. Most of the tumors arising from a single cell have an unlimited proliferative ability yet differ in many other capabilities such as self-renewal, handling stress to avoid death, escaping immune surveillance and invading to form distant metastases. All these differences within tumors are the result of a wide-ranging cellular heterogeneity [4–6]. In addition to heterogeneity, the great level of complexity within tumors is achieved by elaborate interaction of various cell types in response to alteration of the tumor microenvironment [7]. Altogether, cellular plasticity-driven heterogeneity accounts for the development of resistance to chemotherapeutic reagents.

How is tumor cell plasticity achieved? There are basically four fundamental mechanisms, namely mutation, dedifferentiation/transdifferentiation, EMT and inflammation (Fig. 1). Together with epigenetic interactions, tumor cell fusion and exocytic vesicles, these shape the cellular plasticity of the tumors [8]. However, this review focuses only on the fundamental mechanisms that lead to plasticity during the initiation, progression and metastasis of tumors.

The first source of tumor cell plasticity is mutations. As seen in the process of evolution, heterogeneity – variation – must pre-exist in tumor cell populations before any change in the tumor microenvironment occurs. Mutations are key players in cellular heterogeneity within tumors [9]. Driver mutations, such as

chromosomal or microsatellite instability and loss of DNA proofreading or repair machinery, increase tumor cell susceptibility to further mutations to escape cell death or to gain new features required for malignant transformation [10]. Besides, mutations within a solid tumor can give rise to different populations that take part in invasion, metastasis, drug resistance and tumor recurrence [11]. Mutations are not only a component of tumor cell plasticity, but also the basis of all the other mechanisms leading to it. Therefore, every fundamental feature of tumor cell plasticity is based on acquiring sufficient mutations to achieve heterogeneity within the tumors for the growth and expansion of cancer cells.

The second fundamental mechanism that drives tumor cell plasticity is dedifferentiation and/or transdifferentiation, respectively the ability of tumor cells to acquire stem cell-like characteristics supporting continuous proliferation of growing tumors and to shift back to differentiated states [8]. In a given tissue, stem cells are the only cell type that maintains tissue homeostasis following damage or renews the tissue by transforming into any other cell type [8,11,12]. Normally, differentiated cells lose this capability, but differentiated cells within tumors can dedifferentiate, adopting a stem cell-like phenotype [11], and therefore cancer stem cells can arise from the tissue stem cells or from differentiated quiescent cells to sustain tumor growth. This ability is essential to repopulate the cancer stem cell niche against any cellular stress leading to the loss of stem cells [13], thus making stemness and dedifferentiation major assets for tumor cell plasticity.

The third important mechanistic insight into tumor cell plasticity is related to EMT, which grants tumor cells the ability to invade deeper into the tissue or to form distant metastases [14]. When tumors grow to their natural boundaries limited by the surrounding tissues and extracellular matrix, it is a challenge for their metabolically active cells to maintain a nutrient supply sufficient for their high demands [15]. To do so, some tumor cells remodel the extracellular matrix to open up extra space for new tumor cells to invade by coordinating fibroblasts and leukocytes, while others induce angiogenesis to initiate a new blood supply together with endothelial cells, and others still intravasate into the blood circulation to find new places to grow without competition [14,15]. Since only mesenchymal-like cells can activate molecular pathways related to motility, almost all these processes require EMT for cells to gain motility to leave the tumor stroma and form new colonies [16]. However, the mesenchymal state is not as proficient as the epithelial state in terms of proliferative ability [17,18]. Epithelial

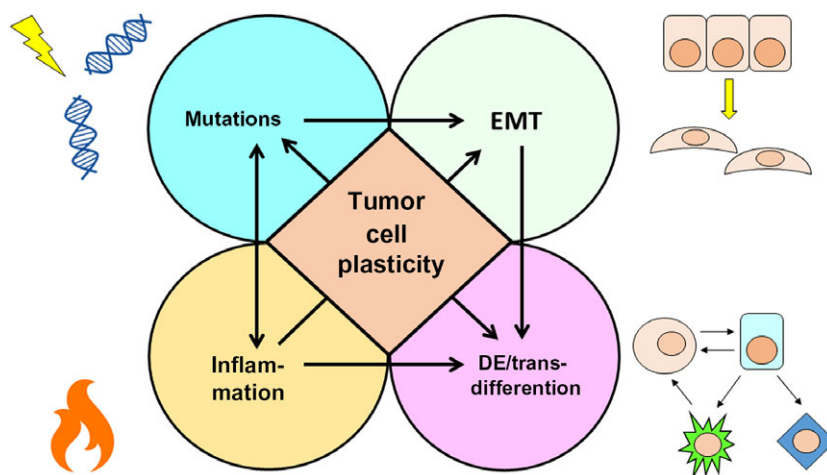


Fig. 1. Mechanisms of tumor cell plasticity. Mutations, EMT, dedifferentiation/transdifferentiation and inflammation are the main mechanisms that give rise to tumor heterogeneity. Heterogeneity is important in enabling tumor cells to adapt to and survive environmental threats.

cells have a basal attachment for efficient proliferation and additionally paracrine growth signals become more effective when the cells are closely packed. Although mesenchymal cells lose these advantages, metastasizing mesenchymal cells still have the capacity to return to a highly proliferative epithelial state to form distant colonies [19–21]. Therefore, epithelial–mesenchymal plasticity is a two-way process helping the tumor cells reprogram their physiology according to the changing demands of the tumor microenvironment.

The last but not least important mechanism of tumor cell plasticity is the ability to create a chronic inflammatory environment within solid tumors. An inflammatory microenvironment can promote tumor growth by triggering the release of growth hormones from surrounding stromal or immune cells, leading to altered cytokine profiles within the tumor, and thus evasion of immunosurveillance. Chronic inflammation also induces a wound healing response, angiogenesis or EMT to enable tumor cells to invade and eventually escape to form metastases [22,23]. Recently, it has been found that a microenvironment that favors growth and increased mutational rates due to reactive oxygen species (ROS) stress and compensatory proliferation upon excessive cell death can initiate tumorigenesis as well [24]. Taken together, an inflammatory environment can be the driving force for tumor cell plasticity. Eventually, the success of tumor cells not only results from their intrinsic ability to alter their own physiology but also depends on how they transform the whole tumor microenvironment to promote tumor growth.

Inflammation as the driving force for tumor cell plasticity

Inflammation is the natural response, elicited through concerted activity of infiltrating leukocytes, cytokine secretion and tissue regeneration, when the host is faced with tissue injury or pathogens, in an attempt to heal a wound or eliminate a pathogen [25]. Normally, inflammation is resolved as soon as the threat to tissue integrity has been eliminated. However, incomplete elimination of a pathogen or failure to properly shut down the inflammatory pathways will result in chronic inflammation. This is one of the hallmarks of cancer, leading to continuous cytokine and protease secretion, and production of ROS and nitric oxides that eventually create an inflammatory microenvironment favoring tumor initiation [24,26]. This microenvironment has many cellular components, namely tumor cells, stromal cells, and innate and adaptive immune cells. It is the intricate signaling among these cell types that tilts the balance towards protumorigenic or antitumorigenic immunity [27].

Among all the molecular pathways that bridge inflammatory regulation of tumor microenvironment and cellular plasticity, nuclear factor κ B (NF- κ B) signaling represents a central hub due to its versatile roles in inflammation and tissue regeneration [28–31]. While chronic inflammation increases susceptibility to cancer development, NF- κ B signaling can also contribute to tumor promotion in cancers not associated with pre-existing inflammation by creating an inflammatory tumor microenvironment [32]. The NF- κ B family comprises proteins that regulate expression of genes

involved in inflammation, innate and adaptive immune responses, cell survival, and cancer [28,33–35]. Although both NF- κ B (e.g. RelA, RelB, cRel, p50, p52) and its regulatory I κ B subunits (e.g. I κ B α , I κ B β , I κ B ϵ , Bcl3, p100, p105) are numerous, little has been revealed of the mutations that affect the functions of these molecules in different diseases [36]. Consequently, our knowledge about their regulation has come mainly from genetic models of upstream regulatory elements, such as IKKs, in cancers of hematopoietic origin [37].

A core element of NF- κ B activation is the IKK complex built from two catalytic subunits, IKK α and IKK β kinases, and a regulatory subunit, NF- κ B essential modulator (NEMO)/IKK γ [29,31]. We know that NF- κ B subunits are regulated by the IKK complex, which is in turn regulated by diverse upstream signaling mechanisms comprising growth, death, stress and pathogen recognition receptors. Activation of NF- κ B transcription factors is highly dependent on both classical and alternative pathways that mediate degradation of I κ B and I κ B-related proteins [29,30]. The classical (or canonical) pathway utilizes IKK β - and IKK γ -dependent I κ B α degradation in response to proinflammatory stimuli (such as interleukin (IL)-1, tumor necrosis factor (TNF)- α and toll-like receptor ligands) releasing active NF- κ B dimers into the nucleus, where they bind to DNA and mediate gene expression [38,39]. In contrast, the alternative (or non-canonical) pathway can be activated by ligands (such as B-cell activating factor (BAFF), lymphotoxin (LT) - $\alpha_1\beta_2$ and CD40L) acting on their specific receptors (BAFFR, LT β R and CD40, respectively) and depends exclusively on IKK α and its upstream kinase, NF- κ B-inducing kinase [40–43]. Throughout the text we will use the term ‘conventional’ to refer to the NF- κ B-dependent (comprising both classical and alternative pathways) functions of IKKs and ‘non-conventional’ for the NF- κ B-independent functions or targets of IKKs. Still, it cannot be ruled out that IKK α/β activity on non-conventional targets or pathways may also be induced through similar upstream elements to those inducing conventional (classical or alternative) NF- κ B signaling pathways [44]; nonetheless, IKK α/β activity on non-conventional targets may not require any of the downstream elements activated by NF- κ B transcription factors [45,46]. To better illustrate conventional and non-conventional targets of IKKs in different cancers, Tables 1 and 2.

Despite the fact that canonical IKKs have structural and functional similarities, they have distinct roles, and diverse phenotypes have been reported for IKK α and IKK β knockout mice [32]. Many studies have addressed the role of IKK subunits in tumor development because of their therapeutic potential, which is

largely absent for NF- κ B or I κ B subunits. Although there is a potential use of specific peptides blocking the interaction of non-kinase subunits of signaling complexes, such as IKK γ in IKK complex, to terminate downstream signaling [47], the number of such studies is still limited compared with studies addressing catalytic subunits of IKK complexes [48–50]. Therefore, in the next section, we concentrate on I κ B kinases as the key mediators of cytokine signaling that are able to orchestrate inflammation in the tumor microenvironment. The focus on both conventional and non-conventional functions, with specific examples of *in vivo* animal models of solid tumors, will explain how these molecules affect tumor cell plasticity by shaping the tumor microenvironment.

Specific roles of IKKs in tumor cell plasticity

Though the roles of canonical NF- κ B signaling have long been well described in many cancers, specific functions of IKK β or IKK α in tumor cell plasticity were poorly characterized until recently. Accumulating evidence suggests that in their non-conventional roles, these kinases possess a greater level of complexity in the way they regulate inflammation, proliferation and metastasis due to their ability to phosphorylate diverse substrates [51]. Table 1 summarizes conventional and non-conventional targets or signaling pathways controlled by IKKs and their effects on tumor cell plasticity in different cancers (also see Fig. 2). Detailed information on the non-conventional IKK α/β targets in different signaling pathways can be found in Table 2. In contrast to IKK β , which spends most of its time in the cytoplasm chasing canonical and non-conventional targets, IKK α translocates into the nucleus where it regulates non-conventional substrates such as p53, histone H3, p27, forkhead box protein A2 (FOXA2) and silencing mediator of retinoic acid and thyroid hormone receptor (SMRT) to direct various mechanisms in inflammation and cancer (Table 2 and Fig. 3) [32,51–53].

IKKs are involved in tumor-elicited inflammation through their indispensable roles in regulating inflammatory cells and their interactions with tumor cells within the tumor microenvironment. Specifically, IKKs regulate proinflammatory or anti-inflammatory signaling machinery through phosphorylation of numerous targets, such as NF- κ B, protein inhibitor of activated STAT1 (PIAS), interferon regulatory factor 3/7 and Tax1 binding protein 1 (TAX1BP1). These targets in turn regulate downstream factors such as TNF, IL-1, IL-6, IL-18, interferon (IFN)- α/β , IFN γ , various other

Table 1. Conventional and non-conventional targets/signaling pathways controlled by IKKs and their effects in tumor cell plasticity in different cancers. Gsk3 β , glycogen synthase kinase 3 β ; iNOS, inducible nitric oxide synthase; NMSC, non-melanoma skin cancer; PFKFB3, 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3; SCC, squamous cell carcinoma.

| Cancer type | IKK subunit | Role | Target molecules/ pathways | Effect in mechanisms regulating tumor cell plasticity | References |
|--------------------------|--------------|------------------|--|--|--------------------------------|
| Colorectal | IKK β | Oncogene | NF- κ B, Wnt, Twist, Bcl-XL, IL-6, STAT3, iNOS, Rac1, NOTCH | Inflammation, stemness, dedifferentiation, metastasis, survival, proliferation | [59,62–65,70,77] |
| | IKK α | Oncogene | NF- κ B, p65, IFN γ , TAX1BP1 | Anti-inflammation, proliferation, tumor tolerance | [54,83,86] |
| Breast | IKK β | Oncogene | FOXO3a, TSC1, GSK3 β , NOTCH, IL-6, Snail, Twist, HER2 | Inflammation, proliferation, survival, stemness | [68,87,88,90–92,94,97,103,104] |
| | IKK α | Oncogene | Myc, mTOR, p27, HER2, Maspin1 | Proliferation, survival, metastasis | [95,98,105–108] |
| Liver | IKK β | Oncogene | NF- κ B, IL-6, STAT3, JNK, SOD, XBP1, PFKFB3 | Inflammation, proliferation, ER stress, survival | [56, 110–113, 116, 123] |
| Skin | IKK α | Oncogene | FOXA2, NOTCH, ATG16 11 | Proliferation, inflammation, survival | [57,117] |
| | IKK β | Tumor suppressor | P16/CDKN2a | Stemness, proliferation | [148] |
| Melanoma, SCC, papilloma | IKK α | Tumor suppressor | ERK, p53, TGF β , cyclin D1, Myc, NPM | Anti-inflammatory, anti-proliferative, stemness, dedifferentiation | [132,134,135,138–141] |
| NMSC | IKK α | Oncogene | EGFR, VEGF-A, Maspin1, MMP9 | Proliferation, EMT, metastasis | [142,143,147] |
| Lung | IKK β | Oncogene | NF- κ B, IL-6, JNK | Proliferation | [150,152] |
| | IKK α | Tumor suppressor | NF- κ B | Anti-inflammatory, Anti-proliferative, tumor surveillance | [153] |
| Prostate | IKK α | Oncogene | Maspin1, IKK β , E2F1 | Metastasis, proliferation, regeneration | [156–158] |
| Ovarian | IKK β | Oncogene | NF- κ B, IL-6, IL-11 | Inflammation, proliferation, survival | [160,161] |
| Pancreas | IKK α | Tumor suppressor | NF- κ B | Anti-inflammatory | [163] |

cytokines and signal transducer and activator of transcription 3 (STAT3) [45,54,55], which also have pleiotropic effects on proliferation, oxidative stress, endoplasmic reticulum (ER) stress and anti-apoptotic machinery, challenging genomic integrity and facilitating mutations, another important mechanism in tumor cell plasticity.

Besides these inflammatory targets, IKKs can also directly or indirectly regulate many proliferative factors, such as (a) cyclin D1, epidermal growth factor receptor (EGFR), tumor growth factor β (TGF β), c-Myc and FOXO3a, which in turn favor mutations due to increased proliferation [46]; (b) many components in survival pathways activated upon stress such as CYLD Lysine 63 Deubiquitinase, X-box-binding protein 1 (XBP1), superoxide dismutase (SOD), Rac Family Small GTPase 1 (Rac1), ATG16 11, receptor-interacting serine/threonine-protein kinase 1 (RIPK1), extracellular signal-regulated kinase (ERK) and c-Jun

N-terminal kinase (JNK) [45,56–59]; and (c) various controlling factors such as p53, Aurora A, ATM Serine/Threonine Kinase 1 or nucleophosmin (NPM) that orchestrate factors to preserve genomic integrity in other cancers [45,51]. IKKs also control factors such as SMRT, FOXA2, histone deacetylase 3, histone H3 α , Smad2/3 or E2F transcription factor 1 (E2F1) participating in NOTCH, Wnt or TGF β pathways [46,51]. The latter is important signaling machinery as highly dividing tumor cells require stemness and dedifferentiation/transdifferentiation to maintain the ability to regenerate or adapt to various stressful conditions within the tumor microenvironment. Last but not least, the specific functions of IKKs in tumor cell plasticity include their abundant functions in controlling EMT through factors such as Wnt, Maspin1, Twist and Snail [8,60]. Formation of tumor cell plasticity highly depends on the regulation of canonical and non-canonical targets by the tissue- and context-specific roles of

Table 2. Detailed information about non-conventional IKK α/β targets in different signaling pathways. The information in this table is compiled mostly from studies with animal tumor models. For more exhaustive information about non-conventional IKK α/β targets in various signaling machinery, not necessarily related to tumor cell plasticity, refer to previous reviews [45,46,51]. AIB1/SRC-3 aliases for NCOA3, nuclear receptor coactivator 3; ER α , estrogen receptor alpha; MMP9, matrix metalloprotease 9; PFKFB3, 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 3.

| Non-conventional IKK targets | Targeted by | NEMO-dependency | Catalytic activity | Cellular localization | Cell type | References |
|------------------------------|--------------------|-----------------|------------------------------------|-----------------------|---|-----------------|
| Histone H3 α | IKK α | Yes | Yes | Nucleus | Fibroblasts, intestinal, breast | [40,53,86,97] |
| SMRT | IKK α | Yes | Yes | Nucleus | Intestinal | [52] |
| NPM | IKK α | No | Yes | Nucleus | Skin | [141] |
| Maspin1 | IKK α | No | No (through p27) | Nucleus | Breast, skin, prostate | [108, 143, 156] |
| p27 | IKK α | No | Yes | Nucleus | Breast (skin, prostate?) | [106] |
| Cyclin D1 | IKK α | No | Yes (through histone H3 α) | Nucleus | Breast | [175] |
| FOXA2 | IKK α | No | Yes | Nucleus | Hepatocytes | [117] |
| E2F1 | IKK α | No | Yes | Nucleus | Prostate | [158] |
| ATG16 I1 | IKK α | Yes | Yes | Cytoplasm | Intestinal | [57] |
| TAX1BP1 | IKK α | No | Yes | Cytoplasm | Fibroblasts | [83] |
| EGFR, VEGF-A, MMP9 | IKK α | Unclear | Unclear | Cytoplasm | Skin | [147] |
| c-Myc, integrin- α 6 | IKK α | No | Yes | Nucleus | Skin | [147] |
| ER α , AIB1/SRC-3 | IKK α | Unclear | Yes | Cytoplasmic | Breast | [97] |
| TSC1 | IKK β | Unclear | Yes | Cytoplasm | Breast, hepatocytes | [88,122] |
| XBP1 | IKK β | Yes | Yes | Cytoplasm | Hepatocytes, (adipocytes?) | [56] |
| PFKFB3 | IKK β | Yes | Yes | Cytoplasm | Breast, fibroblasts | [123] |
| FOXO3a | IKK β | Unclear | Yes | Cytoplasm | Breast | [87] |
| RIPK1 | IKK α/β | Yes | Yes | Cytoplasm | Hepatocytes | [115] |
| NOTCH | IKK α/β | No | No (through SMRT or FOXA2/NUMB) | Nucleus | Intestinal and breast (through SMRT), hepatocytes (through FOXA2) | [52,103] |
| IL-6 | IKK α/β | Yes | No (through NOTCH) | Nucleus | Breast | [103] |
| STAT3 | IKK α/β | Yes | No (through NOTCH) | Nucleus | Breast | [103] |

the IKK kinases. To better illustrate this, we give several examples of the pivotal roles IKKs play in shaping tumor cell plasticity in what follows.

Among many studies associating NF- κ B signaling with almost any cancer in the past two decades, there have been only a limited number providing mechanistic insights, and even fewer describing the roles of IKKs in animal models of tumorigenesis. Since we believe that the complex regulations and interactions within the tumor microenvironment can be best understood through the use of genetic disease models and transgenic animal models, here we select those studies, classified by individual cancers, to demonstrate how canonical or non-canonical IKK functions affect tumor cell plasticity. Of note, however, due to the genetic and physiological differences between these species, there is a possibility that not all the information derived from animal models can be translated into human disease. Recent literature has witnessed an increased interest in patient-derived xenografts or patient-derived organoid models to better approximate and model human diseases. Although these are powerful tools for rapid and

reliable drug screening, toxicological assessment and combinatorial targeting, they fail to establish complex interactions within the tumor microenvironment. To resolve such problems, advances in cell-based co-cultures of multiple origin may be a good alternative. Developments in this area or in related cell-based techniques will eventually eliminate the requirement for animal use in cancer or any other disease research, yet the information derived from such studies about the specific functions of IKKs in various solid malignancies is very limited at the present.

Cancer specific functions of IKKs in tumor cell plasticity

Colorectal cancer

Constitutive NF- κ B activation, a common theme in most cancers, is one of the leading reasons that tumors develop resistance against chemotherapy or radiation. NF- κ B activity is crucial for the activation of anti-apoptotic signaling that protects tumor cells from

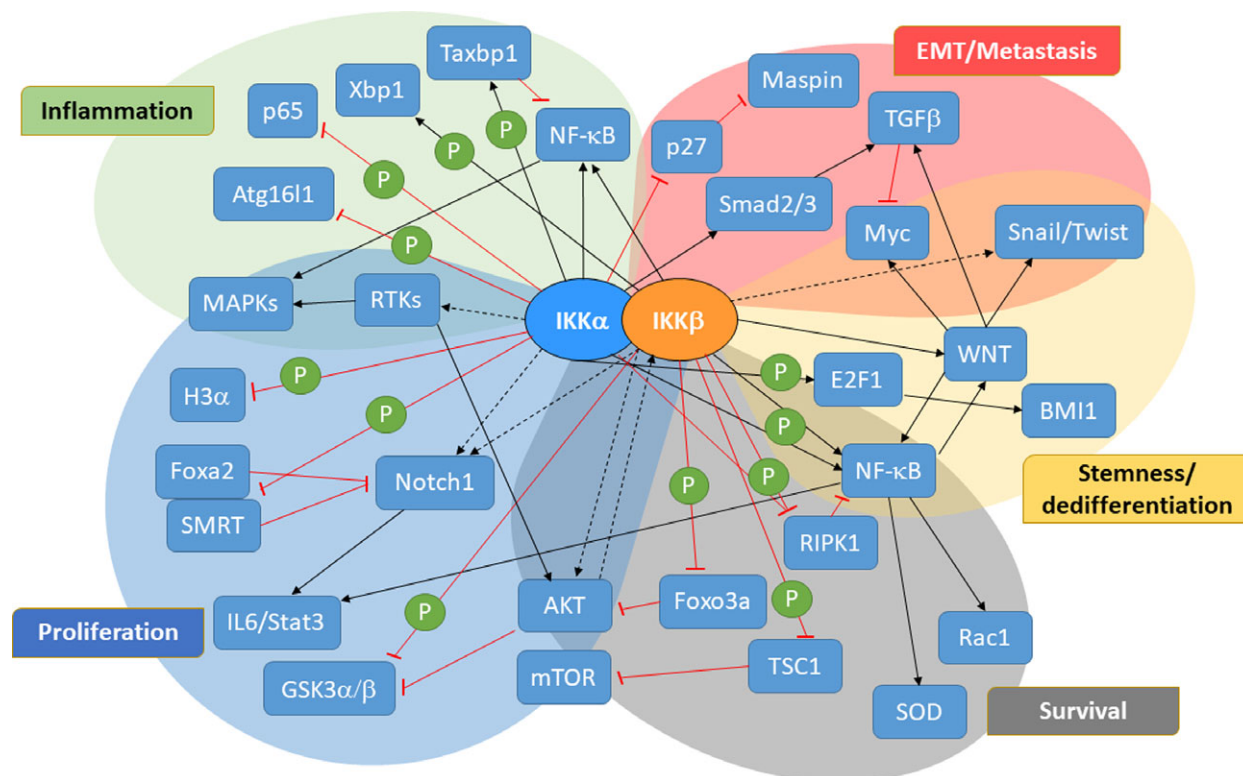


Fig. 2. Signaling pathways governed by IKKs in tumor development and tumor cell plasticity. A summary of the combination of signaling molecules and pathways targeted by IKKs in different cancers. It should be noted that not all pathways are active at the same time or work in the same way in each cancer. Most of these pathways are differentially regulated in different cancers and even in different subtypes. Tables 1 and 2 give more detailed information about IKK target molecules/pathways and how they affect tumor development in various genetic cancer models. GSK3 α/β , glycogen synthase kinase 3 α/β ; H3 α , histone H3 alpha; RTK, receptor tyrosine kinase.

death [61]. Activation of the anti-apoptotic machinery in most cancers, especially in colorectal cancer (CRC), through induction of NF- κ B-dependent gene expression requires IKK β catalytic activity. In a colitis-associated colorectal cancer (CAC) model for tumorigenesis, *Ikk* $\beta^{\Delta IEC}$ (*Ikkbb*) mice were found to have reduced tumor burden due to increased apoptosis through downregulation of Bcl-x_L [62]. On the other hand, *Ikk* $\beta^{\Delta M\Phi}$ mice in a CAC model were found to have reduced tumor number and size due to decreased proliferation resulting from diminished macrophage-derived IL-6 synthesis [62]. Later, it was suggested that macrophage-derived IL-6 and inducible nitric oxide synthase promote tumor growth via inflammatory STAT3 [63,64], activation of which has been shown to regulate proliferation, survival and NF- κ B signaling in a CAC tumor model [65,66].

IKK β -driven inflammation not only provides proliferative or survival advantage but may also influence stemness and differentiation. As suggested in previous studies on different tissues, IKK β -driven inflammation blocks the differentiation of adipocytes, osteoclasts

and vascular endothelia to lock them in a more stem cell-like phenotype through various mechanisms involving regulation of Wnt signaling [67–69]. In our recent work, we have shown that non-stem cells from intestinal epithelia can convert to a stem cell-like phenotype in the presence of constitutive Wnt and NF- κ B activation in spontaneous mouse models of colorectal tumorigenesis [70]. In the same work, activated NF- κ B could induce Wnt signaling or vice versa through the cooperation of their transcription factors at related gene promoters [70]. However, the latter was not mechanistically illustrated and the proof for this mechanism came from a related study in which it was found that both NF- κ B activation and ROS production are controlled by Rac1 in a Wnt-driven colorectal carcinogenesis model by loss of the *Apc* gene [59]. Accordingly, Lgr5⁺ stem cell expansion in Wnt-driven tumors mechanistically required the activation of Rac1 to induce ROS production, and as a result, a marked activation of NF- κ B to protect stem cells from apoptosis [59]. Other studies showed that IKK β expression in cancer-associated fibroblasts (CAFs), also known as

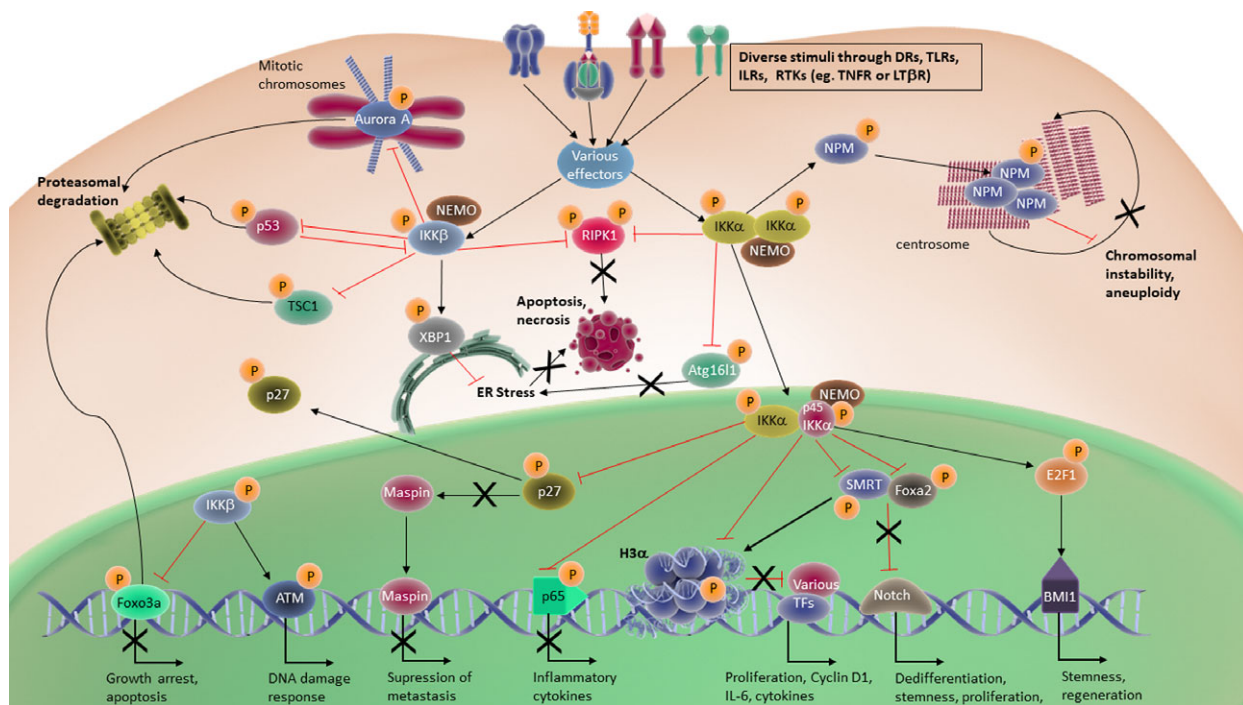


Fig. 3. NF-κB-independent functions of IKKs in tumor development. This figure summarizes some of the most common non-conventional IKK targets in different tumors. Again the regulation on these molecules is determined according to tissue type, tumor stage and other factors within the tumor microenvironment. Therefore, the outcome of any given regulation may show dramatic differences from one cancer to another. Table 2 gives more detailed information about non-conventional IKK targets in various malignancies. ATM1, ATM Serine/Threonine Kinase 1; BMI1, B cell-specific Moloney murine leukemia virus integration site 1; DR, death receptor; IL1R, interleukin 1 receptor; RTK, receptor tyrosine kinase; TF, transcription factor; TLR, toll-like receptor.

intestinal mesenchymal cells, may have potential roles in colorectal tumor cell plasticity [71,72]. Despite the fact that contrasting results were obtained in these studies, the oncogenic potential of IKKβ in CRC may be due to differences in tissue-specific ablation strategies that were constitutive [71] rather than inducible [72]. Although there are some critics requesting further proof on IKKβ-specific roles in CAFs and CRC [73], it is noteworthy that while germline mutations are often compensated during the course of development, studies with inducible models can illustrate better the sporadic nature of cancer.

It remains unknown how IKKβ-driven NF-κB activation triggers the metastatic shift in colorectal tumors. It was first shown that constitutively induced cytokines and a chronic inflammatory phenotype were not sufficient to drive inflammation in the intestine without mitogen-activated protein kinase (MAPK) signaling [74]. MAPKs are required for c-Myc stabilization upon activation of toll-like receptor signaling in Wnt-driven tumor models [75]. Hence, MAPKs and their downstream transcription factors regulate the expression of various cytokines and proliferative factors by co-operating with NF-κB at promoter regions

of target genes [74,76]. However, in later studies, our colleagues have proven that this proinflammatory environment created by constitutive IKKβ activation is not only required for tumor initiation and progression but also for invasion and metastasis upon p53 loss [77]. Consequently, Twist1, an EMT marker, was transcriptionally activated leading to tumor invasion and lymph node metastasis [77].

IKKα regulates intestinal tumor cell plasticity through modulation of inflammation and cell to cell interactions within the tumor microenvironment. It is widely known that immune cells can suppress, tolerate or promote the growth of tumor cells in various cancers, yet the regulation of immune cells in the early stages of tumor development has been poorly described until recently. A recent study on the V12RAS-driven zebrafish melanoma model clearly demonstrated that myeloid cells were recruited to the tumor-initiating cells as soon as the oncogenic transformation occurred [78]. Interaction of both cell types is required for the tumor progression parallel to the wound healing response [78]. Similar to these observations, we have identified a unique intraepithelial localization of myeloid cells in the intestines of mice devoid

of IKK α activation (*Ikkba*^{S181A/S185A} or simply IKK α ^{AA/AA}) upon β -catenin-induced transformation [54]. Mechanistically, IKK α represses an NF- κ B-dependent cytokine expression, required for enhanced recruitment of myeloid cells to the site of tumors [54]. In this model, IKK α -deficient myeloid cells showed M1-like polarization and actively produced IFN γ upon tumorigenesis. Additionally, the tumor suppressive function of myeloid cells in the absence of IKK α activation may not be limited to enhanced paracrine IFN γ secretion, which has been shown to be important in the anti-tumor response [79,80], and they may also phagocytize tumor cells [81] more efficiently, as their specific localization may suggest. Prior to our observations, it was reported that IKK α limits macrophage activation and contributes to the resolution of inflammation via preventing extended promoter binding of RelA and c-Rel [82]. IKK α was found to be crucial in limiting macrophage or dendritic cell activity [82–85]. Given that proinflammatory signaling governed by enhanced NF- κ B activation may exacerbate inflammation in different diseases, IKK α -dependent feedback inhibition may prove to be indispensable for prevention of tissue damage in various pathologies [54]. Previously, it had been reported that nuclear localization of IKK α results in phosphorylation and release of the repressor protein SMRT from NOTCH1 target genes, leading to their aberrant expression upon malignant transformation in CRC [52]. In a recent report, Margalef and colleagues [86] showed that phospho-activated IKK α is first cleaved to a shorter form (p45-IKK α), and then this active and truncated IKK α is translocated to the nucleus where it phosphorylates and release the transcriptional repressor SMRT from the promoters of NOTCH signaling targets, preventing cancer cells from differentiating. The existence of cleaved-nuclear IKK α , a primary form in human patient samples, correlates with tumor stage and prognostic outcome in patients [86]. Thus, IKK α can govern stemness and proliferation of intestinal tumor cells through these non-conventional functions to further enhance their plasticity to tumor microenvironment changes.

Breast cancer

Breast cancer is another example where canonical and non-canonical functions of IKKs affect tumor cell plasticity in various settings related to tumor development and metastasis. One of the first pieces of mechanistic evidence showing that IKK β can regulate non-canonical targets to promote tumorigenesis came from work with animal models of breast cancer [87]. This

study demonstrated a physical FOXO3a–IKK β interaction, followed by FOXO3a phosphorylation, ubiquitination and degradation [87], all of which occur independently of AKT. FOXO3a-depleted tumor cells grow better in mouse xenografts due to extensive NF- κ B activation leading to survival of tumor cells. As a result, FOXO3a-positive breast cancer patients have improved survival due to activation of proapoptotic signaling while FOXO3a[−]IKK β ⁺ patients had worse prognosis [87]. In another study, IKK β was found to connect tumor-elicited inflammation to angiogenesis by directly phosphorylating the mechanistic target of rapamycin (mTOR) repressor TSC complex subunit 1 (TSC1)–TSC2 tumor suppressor complex to initiate angiogenesis within breast tumors [88]. Tumor xenografts with mutant TSC1 lacking a phosphoregulatory site grew poorly, whereas human patients positive for both phospho-TSC1 and phospho-S6K had worse prognosis [88]. Hence, IKK β activation upon hypoxia-induced proinflammatory TNF expression is crucial in angiogenesis induction for further tumor growth and invasion in breast cancer. Additionally, IKK β and NF- κ B were shown to be essential for EMT and metastasis in breast cancer cells [89]. It subsequently became evident that NF- κ B is required for stabilization of Snail or Twist1 in breast cancer cell lines to initiate EMT, invasion and metastasis of breast cancer [90–92]. IKK β was also shown to be crucial for endothelial cell permeability, which is important for the extravasation of metastatic cells [68]. Park and colleagues [93] described that NF- κ B-dependent granulocyte-colony stimulating factor secretion by metastatic breast cancer cells was directing osteoclast differentiation and osteolytic bone metastasis into skeletal tissue, which is frequently observed in breast cancer patients. Similarly, Chen and colleagues [94] observed that IKK β implements a LIN28B/TCF7L2 feedback loop that promotes breast cancer cell stemness and metastasis with the help of *in vivo* xenograft and pharmacological inhibition studies. Briefly, IKK β regulates tumor cell plasticity in breast cancer by regulating many facets of related mechanisms.

In the signaling believed to be involved in breast carcinogenesis, the non-canonical roles of IKK kinases, IKK α in particular, have not been fully investigated until recently. We now know that primary human breast cancer specimens and breast cancer cell lines exhibit aberrant IKK α and IKK β kinase activities that may not involve NF- κ B [95] or its downstream targets [96] to enhance transformation of breast cancer cells. IKK α / β kinases have been also shown to promote mammary tumorigenesis through inhibition of FOXO3a [97] and glycogen synthase kinase 3 β by

the activation of mTORC2, which further regulates the phosphatidylinositol-3-kinase–AKT pathway [98]. Moreover, implication of a role for constitutively active IKK complex and NF- κ B activation in breast cancer cell migration and metastasis has also been reported [99]. Given the important role of NOTCH1 [100,101] and IL-6 signaling in breast tumorigenesis [102], it was recently reported that IKK α and IKK β , independent of NF- κ B, control NOTCH-mediated IL-6/Janus kinase/STAT signaling in breast tumors cells [103]. IKK α was then discovered to be essential for oncogenic transformation in *in vivo* models of Her2/neu-induced breast tumorigenesis [104]. In this study, using two independent mouse models of mammary tumorigenesis, the authors provided evidence supporting the importance of IKK α kinase activity for the increased incidence and advanced onset of ErbB2/Her2-induced breast tumors [104]. This is reinforced with more recent findings as well [105]. In these studies, the enhanced, rather than totally inhibited, tumorigenesis was attributed to the kinase domain of IKK α , which maintains signals essential for the self-renewal potential of tumor-initiating cells. Currently, we know that IKK α regulates the expansion of these cells, in Her2-initiated tumors through a mechanism that involved NF- κ B inducing kinase/IKK α axis induced p27/Kip1 phosphorylation and nuclear exclusion, contributing to augmented tumorigenesis [106]. On the other hand, IKK α was shown to be more efficient than IKK β in activating NF- κ B and the expression of downstream genes involved in cell invasion [107] in Her2-overexpressing breast cancer cells. The prometastatic activity of IKK α was stimulated by receptor activator of NF- κ B (RANK) ligand (RANKL)/RANK to repress Maspin expression [108] and correlated inversely with nuclear p27/Kip1 in human invasive breast carcinoma specimens [106]. The studies summarized above indicate a protumorigenic and prometastatic role of IKK α in the mammary gland in which the kinase activity of IKK α stimulates proliferation of breast cancer cells and protects them from being killed by anti-cancer drugs. Consequently, IKK α can be considered an important regulator of breast cancer progression and a notable target for the development of therapeutic strategies targeting inhibition proliferation, metastasis and drug resistance in breast cancer cells.

Hepatocellular carcinoma

IKK β plays opposing roles in different cells at different time points in hepatic cancer development. Since there were no hepatitis B virus or hepatitis C virus equivalents for generating a hepatitis-induced

carcinogenesis in mice, *Mdr2* knockout mice, which develop cholestatic hepatitis followed by hepatic tumorigenesis, were used in early research [109]. By the use of this model, inflammatory cell-derived TNF is found to promote hepatocyte survival via activation of canonical anti-apoptotic NF- κ B signaling [110]. Interestingly, using an I κ B super-repressor model to silence NF- κ B specifically in hepatocytes, no difference was observed in the tumor development in normal and mutant mice [110]. Only when the same model was used to inactivate NF- κ B conditionally at a later point or when anti-TNF antibody was applied, was tumor growth seriously prevented due to apoptosis induction in the transformed hepatocytes [110]. In sharp contrast to these observations, in a mouse model of chemical carcinogen (diethylnitrosamine; DEN)-induced hepatocellular carcinoma (HCC), it was shown that IKK β deletion in hepatocytes dramatically increased tumor burden while its deletion in macrophages inhibited tumor formation [111]. The observed differences in these two studies may be due to high regenerative abilities of hepatocytes in which IKK β or NF- κ B plays an important protective role against damage-induced cell death and resulting compensatory proliferation upon liver injury by DEN [111]. As shown in another study, NF- κ B activates SOD to counteract ROS-induced stress in hepatocytes [112], whereas in the absence of NF- κ B, ROS induces the activation of JNK and activator protein 1, which eventually leads to hepatocyte death via apoptosis [113]. Further work has clarified that ROS-damage-induced compensatory proliferation in HCC was driven by STAT3 activation in hepatocytes [114]. The contradictory NF- κ B involvement in early and late stages of hepatocellular cancer was further supported by IKK γ /NEMO mutant mice, bearing IKK γ -deficient liver parenchymal cells. These mice spontaneously developed steatosis or multiple tumors in their livers [115]. The absence of IKK γ halts kinase complex formation and causes excessive cell death due to uncontrolled RIPK1 activation. Thus, RIPK1 ablation or RIPK1 kinase inactive knock-in models could prevent hepatocyte apoptosis and HCC in mice [115]. Later studies proposed that the control of RIPK1 to prevent hepatocellular necrosis or apoptosis relies on non-conventional functions of IKKs and differs from their roles in TNF-driven canonical NF- κ B activation-related anti-apoptosis [58]. This observation partially explains the different observations between NF- κ B super-repressor and IKK β -deficient mouse models.

Aside from hepatocytes in HCC, the resident liver macrophages, Kupffer cells, when induced by apoptotic hepatocytes, secrete inflammatory cytokine IL-6, which in turn drives proliferation in hepatocytes through

activation of STAT3 [111,116]. Therefore, when tumor cell plasticity relies rather on myeloid cell-dependent activation of IKK β in HCC, IKK β in myeloid cells does not display a protective role in the late hepatic tumorigenesis as it does in hepatocytes at the earlier stage. In contrast, elevated IKK α levels have been associated with increased oncogenic potential in HCC models of tumorigenesis since IKK α has been found to be activated with elevated TNF expression during hepatitis [117]. Following this, IKK α translocates into the nucleus where it phosphorylates FOXA2 to suppress its downstream targets such as NUMB which in turn is a NOTCH1, an important transcription factor that regulates cell fate and differentiation during various developmental processes, inhibitor [117]. Therefore, this specific role for IKK α helps the proliferation of hepatocytes in an inflammation-driven and NOTCH1-dependent manner [117].

IKK α/β activation in HCC can be also induced through other pathways than tumor necrosis factor receptor (TNFR) signaling. Indeed, more recent studies with hepatitis-driven cancer models indicated that TNFR can be dispensable for NF- κ B activation and its downstream effects in chronic hepatitis-driven HCC [118–120]. Upon observing that LTs and their receptor LT β R were highly upregulated in HCC, Haybaeck and colleagues [118] developed a chronic HCC mouse model (to simulate hepatitis B virus- and hepatitis C virus-driven chronic hepatitis and HCC) with LT-overexpressing hepatocytes. They showed that only an LT (α and β) response of hepatocytes leads to IKK α/β activation in tumor cells. Therefore, only LT β R activation is required for oncogenic transformation upon chronic hepatitis while TNFR1 was found to be dispensable. Later studies, with mouse models of high-fat-diet-induced steatohepatitis-driven HCC, showed that LTs or LIGHT (TNFSF14) produced by natural killer T cells or CD8⁺ cells specifically signaled through LT β R on hepatocytes to induce steatohepatitis and hepatocyte damage through the activation of the canonical NF- κ B pathway for hepatocellular tumorigenesis [119,120]. Hence, the differences in the observed results are probably due to differences in experimental models. More satisfactory conclusions about the upstream factors leading to IKK activation in hepatocellular tumorigenesis may be derived from advanced mouse models that better mimic the development of human disease.

As with other mechanisms, ER stress can alter the tumor microenvironment and tumor cell plasticity through modulation of immune responses, fueling tumor development [121]. Earlier studies proposed that IKK β control on TSC1 and the mTOR signaling

pathway may regulate glucose metabolism and insulin resistance through the regulation of insulin receptor substrate 1 in hepatocytes [122]. Similarly IKK β has been shown to regulate glutamine deprivation, resulting from increased anabolic activity in highly dividing cancer cells, through its direct control on 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase isoform 3 in the glycolytic pathway [123]. Recent studies have provided solid connections between ER stress, inflammation and cancer in certain malignancies [124]. Especially in liver and colorectal cancers, elements of ER stress, nutrition and metabolism have been shown to be connected with tumor-elicited inflammation in tumor suppression and maintenance of the stem cell niche [125,126]. The important actors in ER stress, such as XBP1 and ATG16L1, have been shown to be associated with IKK α/β in the modulation of inflammation through protection from ER stress-induced apoptosis [57] and glucose homeostasis through their interaction [56]. All these observations indicate that IKKs can be the key regulators of ER stress- and tumor-elicited inflammation. Therefore, better understanding of IKK kinases in the mechanism of ER stress and cancer development is required to develop better treatment options for these cancers.

Skin cancers

IKK α has emerged as an innate surveillant in the skin controlling homeostasis and suppressing tumor formation. Mice with IKK α deletion exhibit numerous morphological abnormalities and die soon after birth, displaying defects in keratinocyte terminal differentiation and epidermal hyperplasia [127–129]. Reintroduction of an IKK α transgene in IKK α -deficient keratinocytes in mice induces terminal differentiation and rescues the phenotype of *Ikk α ^{-/-}* (*Ikbka*) mice [130–132]. This function of IKK α in the formation of epidermis was independent of the kinase domain and NF- κ B activities [130,131,133]. Decreased IKK α expression has been reported in several mouse [132–137] and human squamous cell carcinomas (SCCs) [133–136] suggesting that IKK α has a key role as a tumor suppressor in the skin. It has been proposed that IKK α expression level is important for skin tumor development. Liu *et al.* [132] highlighted the importance of IKK α in the development of squamous cell carcinoma in mice and humans. They showed that overexpression of an IKK α transgene in the epidermis (Lori-IKK α -Tg mice) enhanced terminal differentiation and inhibited carcinoma development and tumor metastasis in a 7,12-dimethylbenzanthracene (DMBA) and 12-*O*-tetradecanoylphorbol-13-acetate (TPA)

chemical model of skin carcinogenesis [132]. In the same study they identified novel IKK α mutations in human SCC samples, suggesting that the aggressiveness of SCC is inversely correlated with IKK α expression [132]. Moreover, elevated IKK α expression suppressed epidermal thickness and skin inflammation in a dose-dependent manner. In a follow-up study, the authors used two transgenic mouse lines with different expression levels of IKK α in the basal epidermis and demonstrated that higher IKK α levels are critical for mouse embryonic development and suppression of excessive keratinocyte mitosis [137]. Also, in DMBA/TPA-induced skin carcinogenesis IKK α hemizygotes developed more papilloma and carcinoma than the control group. Most of the carcinoma and some papilloma of *Ikk α ^{+/-}* mice displayed reduced IKK α expression [138]. Furthermore, *Ikk α* deletion in basal epidermal keratinocytes (using keratin promoter 5) caused the same phenotype observed in *Ikk α ^{-/-}* mice and was reversed by introduction of a keratinocyte-specific IKK α transgene [139]. Therefore, during SCC development the levels of IKK α are critically reduced, yet a sufficient dose is required for skin development and protection. In UVB-triggered carcinogenesis, IKK α reduction induced increased ERK activity and proliferation in the skin resulting in enhanced tumor numbers and p53 mutations [140]. Lastly, other studies suggested that inflammation-activated IKK α helps to protect skin cells against genomic instability by phospho-activation (at Ser125) of NPM oligomerization, activity of which blocks centrosome doubling [141]. As a result, in human SCC samples both IKK α and NPM are often observed to be decreased. Therefore, IKK α and NPM have profound effects in preventing aneuploidy and protecting genomic integrity in squamous skin epithelia [141].

In contrast, implication of IKK α in the malignant potential of skin tumors in the DMBA/TPA model has also been reported [142]. Although the essential role of IKK α in keratinocyte differentiation and stratification has been confirmed, an IKK α -overexpressing three-dimensional human skin equivalent (HaCaT and fibroblast cultures) has shown marked increase in keratinocyte proliferation and invasiveness [143]. In a chemically induced skin carcinogenesis model initiated by DMBA/TPA [144–146], IKK α reduced ERK activation affecting carcinoma development and vascular endothelial growth factor A (VEGF-A) expression impeding tumor invasion, independent of kinase activity and NF- κ B activation [132,138]. Similarly, in non-melanoma skin cancer (NMSC) IKK α has shown protumorigenic roles depending on its subcellular localization [147]. NMSC mouse models were

developed by the use of Tg.AC mice, expressing *v-Ha-Ras* oncogene, and these mice were mated to cytoplasmic or nuclear expressing human-IKK α transgenic mice. As a result, cytoplasmic IKK α -induced tumorigenesis was dependent on EGFR, VEGF-A and matrix metalloprotease 9, while nuclear IKK α transgene-induced oncogenicity was dependent on c-Myc, Maspin and integrin- α 6 expression [147]. Consequently, IKK α exerts its oncogenicity in NMSCs while acting as a tumor suppressor in melanoma and papilloma, which further demonstrates its versatility in tumor cell plasticity, depending both on cell type and cellular localization. In contrast to numerous studies revealing the role IKK α , there are only a few associating IKK β with skin tumorigenesis. A recent work showed IKK β to act as a tumor suppressor in basal skin cancer models through regulation of p16-Ink4a (CDKN2a) [148]. The authors argued that IKK β mediates resistance towards tumor development in epidermal cells but not in other cell types. Therefore, both IKK β and IKK α may play a dual role in skin cancer depending on the cell type and interactions within the tumor microenvironment.

Lung, prostate and other cancers

Nicotine-derived nitrosamine ketone, a tobacco smoke byproduct, is a chemical carcinogen frequently used to generate lung cancer models in rodents. Although it was reported to induce inflammation and proliferation in bronchial epithelial cells through activation of NF- κ B and cyclin D1 [149], previous studies using these models failed to report an association of tobacco smoke with increased tumor progression *in vivo*. However, a modified regime of tobacco exposure has been recently utilized to prevent desensitization of pulmonary cells to tobacco smoke [150] in a K-Ras (K-RasG12D)-driven pulmonary carcinogenesis model [151]. It indicated that chronic inflammation due to repeated tobacco smoke exposure promoted tumor growth in an IKK β - and JNK-dependent manner. Consequently, tobacco smoke-driven chronic inflammation utilizes IKK β for the activation of inflammatory signaling in macrophages leading to tissue damage and, following MAPK activation, to promote tumor growth via increased proliferation [150]. Later IL-6, a target of NF- κ B activation in myeloid cells, was found to be a mediator of increased proliferation in inflammation-driven lung cancer [152].

The role of IKK α in macrophage regulation of lung tumorigenesis strikingly differs from that of colorectal cancer. These differences became more apparent in a study where IKK α kinase dead (KD) mutant animals

were generated [153] instead of the use of K-RasG12D-driven lung adenocarcinoma model. These mice produced sporadic lung tumors resembling human small cell carcinoma, marked by the upregulation of markers such as K5, p63 and Trim29 [153]. Reintroduction of transgenic K5 and wild-type bone marrow transplantation reversed the SCC-like phenotype in IKK α -KD mice [153]. Therefore, IKK α downregulation in bronchial epithelia triggers an inflammatory response that causes polarization of macrophages towards the tumor-promoting M2 phenotype. While both in colon and lung cancers IKK α plays an anti-inflammatory role to augment activation of NF- κ B-driven inflammation, in lung cancer the absence of IKK α leads to excessive inflammation that arguably involves T-cell activation. As a result, the latter eventually leads to the recruitment of anti-inflammatory M2 macrophages into lung tumors.

Although most prostate cancers (CaP) are androgen-dependent and responds quite efficiently to androgen ablation, they become androgen-independent later on [154]. Therefore, the major risk for CaP patients is the development of drug resistance. While tumor initiation and progression clearly involve inflammatory processes, such as NF- κ B and STAT3 activation, the role of inflammation in metastatic progression still awaits more evidence. In one study, IKK α was found to be an important mediator of CaP metastasis in a mouse model of prostate adenocarcinoma (transgenic adenocarcinoma of the mouse prostate, in which the SV40 large T antigen is specifically expressed in the prostate epithelium) that closely reflects the pathogenesis of human prostate cancer [155]. Using these mice, Luo *et al.* [156], demonstrated that IKK α kinase activity regulates the interplay between inflammation and prostate metastatogenesis. They suggested that IKK α activation and nuclear translocation are the critical events at the onset of metastasis since it is required for the repression of the metastasis suppressor Maspin. IKK α activation relies on the interaction of myeloid and lymphoid cells that are attracted into the tumors and secreted cytokines such as RANKL and LT [156]. The activation of IKK α in CaP cells was shown to be IKK β -dependent but NF- κ B-independent [156,157]. Leukocytes and B cells displaying IKK β activation were recruited to the vanishing tumor and produced cytokines, such as LT, which in turn activated IKK α on the surviving CaP cells resulting in increased metastasis [157]. However, the effects of infiltrating B cells on IKK α activation were not limited to the metastasis but also involved in castration resistance and tumor recurrence in CaP through the regeneration of tumor cells [158]. In IKK α -deficient (IKK $\alpha^{AA/AA}$) CaP mice,

tumor recurrence was considerably diminished. Mechanistically, upon activation by B-cell-derived LTs in CaP tumors, IKK α was localized to the nucleus where it associated with transcription factor E2F1 on the *Bmi1* promoter to activate a cancer stem cell renewal pathway [158].

In other cancers, such as ovarian and pancreatic, it has also been reported that myeloid-derived IL-6 is important in inflammation-induced tumorigenesis models through activation of STAT3 [159–161]. Therefore, IKK β mostly, but not necessarily, exerts its pro-tumorigenic function through canonical NF- κ B signaling via activation of proinflammatory cytokine signaling in myeloid cells or anti-apoptotic pathways to protect tumor cells from stress-induced cell death. Indeed, proinflammatory cytokines such as TNF, IL-6, IL-11 and IL-22 activate proliferative STAT3 signaling that converges to canonical NF- κ B signaling to orchestrate diverse mechanisms leading to tumor cell plasticity in solid tumors [37]. IKK α has tumor suppressive roles through suppression of destructive inflammation in skin, lung [139,153,162] and possibly in pancreatic cancer [163], but the recent literature has shown oncogenic roles of IKK α in driving tumor initiation, progression and metastasis in various other cancers [54,106,117,156]. The differential involvement of IKK α in various cancers can depend on upstream signals in a spatiotemporal manner and on variations in tumor microenvironment such as the microbiota or the cytokine milieu in different organs [54,164].

Perspectives

Since NF- κ B signaling is constitutively activated in numerous cancers including hematological and solid malignancies, the pharmaceutical industry has aggressively sought to develop therapeutic strategies to target the NF- κ B pathway [37]. There are as yet many constraints in developing effective IKK inhibitors that could make an ideal drug. They should (a) be very specific to NF- κ B signaling (but not to other signaling pathways); (b) selectively target specific NF- κ B components in a given disease, (c) be more effective on tumor cells than on any other cells; (d) have an effect that is transient and highly reversible to prevent harmful side effects on innate immunity and extended immunosuppression; and (e) have efficacy, dose, delivery method and delivery schedules that are carefully determined for a single agent or in combination with others [165]. With much effort put in by pharmaceutical companies, inhibitors targeting NF- κ B, NEMO and IKK β have been developed [165]. Early success with these was reached in a wide array of preclinical studies in

hematological malignancies, such as multiple myeloma, and some other solid malignancies where tumor development was largely dependent on NF- κ B signaling [37]. However, none of this preclinical knowledge could be translated into clinical outcomes and none of these inhibitors could get clinical approval due to toxicity linked with systemic NF- κ B suppression [166]. Due to the complexity of NF- κ B regulation in various cells within the tumor microenvironment, systemic inhibition of any of the I κ B kinases would lead to potential development of septic shock, as seen in many different animal cancer models [82,167,168]. Even indirect inhibitors, such as the proteasomal inhibitor bortezomib that targets multiple signaling pathways, could not avoid dose-limiting toxicities as their efficacy was significantly limited due to their interference with systemic NF- κ B activities [169].

Although therapeutic approaches targeting the core elements of NF- κ B signaling, such as IKKs, in principle have the potential to block cancer-promoting activity, they would also challenge the essential functions of this pathway in immunity, inflammation and anti-apoptosis [165]. It gets even more complicated when the effects of NF- κ B signaling, being vastly tissue- and context-specific, have to be considered. That is the reason that many new studies are carried out with more specific strategies to only intervene with the NF- κ B pathway in a cancer- and tissue-selective manner without affecting NF- κ B's essential functions in other tissues [166]. Such approaches seem promising and indispensable to preserve non-conventional functions of specific IKK subunits. However, the biggest challenge is still how to develop IKK α - or IKK β -specific inhibitors due to the great similarity in their catalytic domains. Over the past two decades, no such inhibitors have been developed [37] by the pharmaceutical industry.

Regardless of the pitfalls in developing effective targeting of the NF- κ B pathway, the studies we presented in this review on IKK α and IKK β involvement in tumor cell plasticity still support the idea that these proteins are potential therapeutic targets for many different malignancies. There are yet challenges in putting the theoretical knowledge into practice because of tumor cell plasticity. Problems include recurrence of tumors due to acquired resistance, and non-cancer stem cells gaining a stem cell-like phenotype [170]. Given that heterogeneity stemming from tumor cell plasticity makes the latter harder to control, the direct killing of cells by chemotherapeutic agents may lead to greater tumor stem cell propagation as a side effect [171]. Moreover, tumor cells cannot be targeted by one or more drugs efficiently due to coexistence of different subpopulations of them within tumors. On the

other hand, intervention on EMT, as seen with many drugs, seems to increase colonization efficiency of circulating metastatic cells [20]. Due to the aforementioned obstacles, therapies aiming at immune cells within the tumor microenvironment may reverse drug resistance. We suggest that it could be essential for the future of clinical cancer research, such as reprogramming blood cells for the initiation of anti-tumor activity [172] or cell-specific inactivation of inflammatory molecules like IKKs by means of gene therapy. In particular, to circumvent associated problems with the development of a specific inhibitor, one may propose the design of non-ATP competitive kinase inhibitors (for example small molecule inhibitors), which may offer better specificity and prevention of off-target effects [173,174]. For the design of such inhibitors, differences in structural information on catalytic domains of individual IKKs or structural differences in phosphorylation or docking sites of non-overlapping substrates targeted by these kinases can be used. Such information will be instrumental in identifying novel non-canonical targets. Since we have pointed out here that inflammatory pathways are the very source of plasticity within tumors, it is important to have a better understanding of regulatory pathways in inflammation-driven tumorigenesis to develop more advanced strategies to tackle cancer.

Acknowledgements

SIG is supported by BİDEB 2232, TÜBA-GEBİP and BAGEP fellowships.

Conflict of interest

The authors declare no conflict of interest.

Author contributions

SIG and MAD drafted the manuscript. TLC checked the manuscript and made critical changes and contributions. All authors read and approved the final manuscript.

References

- 1 Balkwill F & Mantovani A (2001) Inflammation and cancer: back to Virchow? *Lancet* **357**, 539–545.
- 2 Parkin DM (2006) The global health burden of infection-associated cancers in the year 2002. *Int J Cancer* **118**, 3030–3044.
- 3 Mantovani A, Allavena P, Sica A & Balkwill F (2008) Cancer-related inflammation. *Nature* **454**, 436–444.

- 4 Marusyk A, Almendro V & Polyak K (2012) Intra-tumour heterogeneity: a looking glass for cancer? *Nat Rev Cancer* **12**, 323–334.
- 5 Clevers H (2011) The cancer stem cell: premises, promises and challenges. *Nat Med* **17**, 313–319.
- 6 Magee JA, Piskounova E & Morrison SJ (2012) Cancer stem cells: impact, heterogeneity, and uncertainty. *Cancer Cell* **21**, 283–296.
- 7 Schwitalla S (2014) Tumor cell plasticity: the challenge to catch a moving target. *J Gastroenterol* **49**, 618–627.
- 8 Varga J, De Oliveira T & Greten FR (2014) The architect who never sleeps: tumor-induced plasticity. *FEBS Lett* **588**, 2422–2427.
- 9 Meacham CE & Morrison SJ (2013) Tumour heterogeneity and cancer cell plasticity. *Nature* **501**, 328–337.
- 10 Fodde R, Smits R & Clevers H (2001) APC, signal transduction and genetic instability in colorectal cancer. *Nat Rev Cancer* **1**, 55–67.
- 11 Beck B & Blanpain C (2013) Unravelling cancer stem cell potential. *Nat Rev Cancer* **13**, 727–738.
- 12 Welling M & Geijsen N (2013) Uncovering the true identity of naive pluripotent stem cells. *Trends Cell Biol* **23**, 442–448.
- 13 Shoshani O & Zipori D (2015) Stress as a fundamental theme in cell plasticity. *Biochim Biophys Acta* **1849**, 371–377.
- 14 Kalluri R & Weinberg RA (2009) The basics of epithelial-mesenchymal transition. *J Clin Invest* **119**, 1420–1428.
- 15 Kalluri R (2009) EMT: when epithelial cells decide to become mesenchymal-like cells. *J Clin Invest* **119**, 1417–1419.
- 16 Friedl P & Alexander S (2011) Cancer invasion and the microenvironment: plasticity and reciprocity. *Cell* **147**, 992–1009.
- 17 Vega S, Morales AV, Ocana OH, Valdes F, Fabregat I & Nieto MA (2004) Snail blocks the cell cycle and confers resistance to cell death. *Genes Dev* **18**, 1131–1143.
- 18 Mejlvang J, Kriaievska M, Vandewalle C, Chernova T, Sayan AE, Berx G, Mellon JK & Tulchinsky E (2007) Direct repression of cyclin D1 by SIP1 attenuates cell cycle progression in cells undergoing an epithelial mesenchymal transition. *Mol Biol Cell* **18**, 4615–4624.
- 19 Gao D, Joshi N, Choi H, Ryu S, Hahn M, Catena R, Sadik H, Argani P, Wagner P, Vahdat LT *et al.* (2012) Myeloid progenitor cells in the premetastatic lung promote metastases by inducing mesenchymal to epithelial transition. *Cancer Res* **72**, 1384–1394.
- 20 Ocana OH, Corcoles R, Fabra A, Moreno-Bueno G, Acloque H, Vega S, Barrallo-Gimeno A, Cano A & Nieto MA (2012) Metastatic colonization requires the repression of the epithelial-mesenchymal transition inducer Prrx1. *Cancer Cell* **22**, 709–724.
- 21 Polyak K & Weinberg RA (2009) Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer* **9**, 265–273.
- 22 Colotta F, Allavena P, Sica A, Garlanda C & Mantovani A (2009) Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* **30**, 1073–1081.
- 23 Hanahan D & Coussens LM (2012) Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell* **21**, 309–322.
- 24 Grivennikov SI, Greten FR & Karin M (2010) Immunity, inflammation, and cancer. *Cell* **140**, 883–899.
- 25 Coussens LM & Werb Z (2002) Inflammation and cancer. *Nature* **420**, 860–867.
- 26 Hanahan D & Weinberg RA (2011) Hallmarks of cancer: the next generation. *Cell* **144**, 646–674.
- 27 Lin WW & Karin M (2007) A cytokine-mediated link between innate immunity, inflammation, and cancer. *J Clin Invest* **117**, 1175–1183.
- 28 Hayden MS & Ghosh S (2011) NF-kappaB in immunobiology. *Cell Res* **21**, 223–244.
- 29 Solt LA & May MJ (2008) The I κ B kinase complex: master regulator of NF-kappaB signaling. *Immunol Res* **42**, 3–18.
- 30 Beinke S & Ley SC (2004) Functions of NF-kappaB1 and NF-kappaB2 in immune cell biology. *Biochem J* **382**, 393–409.
- 31 Karin M & Ben-Neriah Y (2000) Phosphorylation meets ubiquitination: the control of NF-[kappa]B activity. *Annu Rev Immunol* **18**, 621–663.
- 32 Liu F, Xia Y, Parker AS & Verma IM (2012) IKK biology. *Immunol Rev* **246**, 239–253.
- 33 Bonizzi G & Karin M (2004) The two NF-kappaB activation pathways and their role in innate and adaptive immunity. *Trends Immunol* **25**, 280–288.
- 34 Hu MC & Hung MC (2005) Role of I κ B kinase in tumorigenesis. *Future Oncol* **1**, 67–78.
- 35 Ghosh S & Hayden MS (2008) New regulators of NF-kappaB in inflammation. *Nat Rev Immunol* **8**, 837–848.
- 36 Hacker H & Karin M (2006) Regulation and function of IKK and IKK-related kinases. *Sci STKE* **2006**, re13.
- 37 DiDonato JA, Mercurio F & Karin M (2012) NF-kappaB and the link between inflammation and cancer. *Immunol Rev* **246**, 379–400.
- 38 Li ZW, Chu W, Hu Y, Delhase M, Deerincq T, Ellisman M, Johnson R & Karin M (1999) The IKKbeta subunit of I κ B kinase (IKK) is essential for nuclear factor kappaB activation and prevention of apoptosis. *J Exp Med* **189**, 1839–1845.

- 39 Ghosh S & Karin M (2002) Missing pieces in the NF-kappaB puzzle. *Cell* **109** (Suppl), S81–S96.
- 40 Park GY, Wang X, Hu N, Pedchenko TV, Blackwell TS & Christman JW (2006) NIK is involved in nucleosomal regulation by enhancing histone H3 phosphorylation by IKKalpha. *J Biol Chem* **281**, 18684–18690.
- 41 Senftleben U, Cao Y, Xiao G, Greten FR, Krahn G, Bonizzi G, Chen Y, Hu Y, Fong A, Sun SC *et al.* (2001) Activation by IKKalpha of a second, evolutionary conserved, NF-kappa B signaling pathway. *Science* **293**, 1495–1499.
- 42 Sun SC (2012) The noncanonical NF-kappaB pathway. *Immunol Rev* **246**, 125–140.
- 43 Sun SC (2017) The non-canonical NF-kappaB pathway in immunity and inflammation. *Nat Rev Immunol* **17**, 545–558.
- 44 Hinz M & Scheidereit C (2014) The I κ B kinase complex in NF- κ B regulation and beyond. *EMBO Rep* **15**, 46–61.
- 45 Chariot A (2009) The NF-kappaB-independent functions of IKK subunits in immunity and cancer. *Trends Cell Biol* **19**, 404–413.
- 46 Colomer C, Marruecos L, Vert A, Bigas A & Espinosa L (2017) NF-kappaB members left home: NF-kappaB-independent roles in cancer. *Biomedicines* **5**, E26.
- 47 May MJ, D'Acquisto F, Madge LA, Glockner J, Pober JS & Ghosh S (2000) Selective inhibition of NF-kappaB activation by a peptide that blocks the interaction of NEMO with the IkappaB kinase complex. *Science* **289**, 1550–1554.
- 48 Carvalho G, Fabre C, Braun T, Grosjean J, Ades L, Agou F, Tasdemir E, Boehrer S, Israel A, Veron M *et al.* (2007) Inhibition of NEMO, the regulatory subunit of the IKK complex, induces apoptosis in high-risk myelodysplastic syndrome and acute myeloid leukemia. *Oncogene* **26**, 2299–2307.
- 49 Solt LA, Madge LA & May MJ (2009) NEMO-binding domains of both IKKalpha and IKKbeta regulate IkappaB kinase complex assembly and classical NF-kappaB activation. *J Biol Chem* **284**, 27596–27608.
- 50 Tian F, Zhou P, Kang W, Luo L, Fan X, Yan J & Liang H (2015) The small-molecule inhibitor selectivity between IKKalpha and IKKbeta kinases in NF-kappaB signaling pathway. *J Recept Signal Transduct Res* **35**, 307–318.
- 51 Espinosa L, Margalef P & Bigas A (2015) Non-conventional functions for NF-kappaB members: the dark side of NF-kappaB. *Oncogene* **34**, 2279–2287.
- 52 Fernandez-Majada V, Aguilera C, Villanueva A, Vilardell F, Robert-Moreno A, Aytes A, Real FX, Capella G, Mayo MW, Espinosa L *et al.* (2007) Nuclear IKK activity leads to dysregulated notch-dependent gene expression in colorectal cancer. *Proc Natl Acad Sci USA* **104**, 276–281.
- 53 Yamamoto Y, Verma UN, Prajapati S, Kwak YT & Gaynor RB (2003) Histone H3 phosphorylation by IKK-alpha is critical for cytokine-induced gene expression. *Nature* **423**, 655–659.
- 54 Goktna SI, Canli O, Bollrath J, Fingerle AA, Horst D, Diamanti MA, Pallangyo C, Bennecke M, Nebelsiek T, Mankan AK *et al.* (2014) IKKalpha promotes intestinal tumorigenesis by limiting recruitment of M1-like polarized myeloid cells. *Cell Rep* **7**, 1914–1925.
- 55 Grivennikov SI & Karin M (2010) Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev* **21**, 11–19.
- 56 Liu J, Ibi D, Taniguchi K, Lee J, Herrema H, Akosman B, Mucka P, Salazar Hernandez MA, Uyar MF, Park SW *et al.* (2016) Inflammation improves glucose homeostasis through IKKbeta-XBP1s interaction. *Cell* **167**, 1052–1066. e18
- 57 Diamanti MA, Gupta J, Bennecke M, De Oliveira T, Ramakrishnan M, Braczynski AK, Richter B, Beli P, Hu Y, Saleh M *et al.* (2017) IKKalpha controls ATG16L1 degradation to prevent ER stress during inflammation. *J Exp Med* **214**, 423–437.
- 58 Dondelinger Y, Jouan-Lanhout S, Divert T, Theatre E, Bertin J, Gough PJ, Giansanti P, Heck AJ, Dejardin E, Vandenabeele P *et al.* (2015) NF-kappaB-independent role of IKKalpha/IKKbeta in preventing RIPK1 kinase-dependent apoptotic and necroptotic cell death during TNF signaling. *Mol Cell* **60**, 63–76.
- 59 Myant KB, Cammareri P, McGhee EJ, Ridgway RA, Huels DJ, Cordero JB, Schwitalla S, Kalna G, Ogg EL, Athineos D *et al.* (2013) ROS production and NF-kappaB activation triggered by RAC1 facilitate WNT-driven intestinal stem cell proliferation and colorectal cancer initiation. *Cell Stem Cell* **12**, 761–773.
- 60 Affara NI & Coussens LM (2007) IKKalpha at the crossroads of inflammation and metastasis. *Cell* **129**, 25–26.
- 61 Karin M (2008) The IkappaB kinase - a bridge between inflammation and cancer. *Cell Res* **18**, 334–342.
- 62 Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF & Karin M (2004) IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* **118**, 285–296.
- 63 Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, Scheller J, Rose-John S, Cheroutre H, Eckmann L *et al.* (2009) IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell* **15**, 103–113.

- 64 Shaked H, Hofseth LJ, Chumanevich A, Chumanevich AA, Wang J, Wang Y, Taniguchi K, Guma M, Shenouda S, Clevers H *et al.* (2012) Chronic epithelial NF-kappaB activation accelerates APC loss and intestinal tumor initiation through iNOS up-regulation. *Proc Natl Acad Sci USA* **109**, 14007–14012.
- 65 Bollrath J, Phesse TJ, von Burstin VA, Putoczki T, Bennecke M, Bateman T, Nebelsiek T, Lundgren-May T, Canli O, Schwitalla S *et al.* (2009) gp130-mediated Stat3 activation in enterocytes regulates cell survival and cell-cycle progression during colitis-associated tumorigenesis. *Cancer Cell* **15**, 91–102.
- 66 Lee H, Herrmann A, Deng JH, Kujawski M, Niu G, Li Z, Forman S, Jove R, Pardoll DM & Yu H (2009) Persistently activated Stat3 maintains constitutive NF-kappaB activity in tumors. *Cancer Cell* **15**, 283–293.
- 67 Chang J, Liu F, Lee M, Wu B, Ting K, Zara JN, Soo C, Al Hezaimi K, Zou W, Chen X *et al.* (2013) NF-kappaB inhibits osteogenic differentiation of mesenchymal stem cells by promoting beta-catenin degradation. *Proc Natl Acad Sci USA* **110**, 9469–9474.
- 68 Ashida N, Senbanerjee S, Kodama S, Foo SY, Coggins M, Spencer JA, Zamiri P, Shen D, Li L, Sciuto T *et al.* (2011) IKKbeta regulates essential functions of the vascular endothelium through kinase-dependent and -independent pathways. *Nat Commun* **2**, 318.
- 69 Sui Y, Park SH, Xu J, Monette S, Helsley RN, Han SS & Zhou C (2014) IKKbeta links vascular inflammation to obesity and atherosclerosis. *J Exp Med* **211**, 869–886.
- 70 Schwitalla S, Fingerle AA, Cammareri P, Nebelsiek T, Goktuna SI, Ziegler PK, Canli O, Heijmans J, Huels DJ, Moreaux G *et al.* (2013) Intestinal tumorigenesis initiated by dedifferentiation and acquisition of stem-cell-like properties. *Cell* **152**, 25–38.
- 71 Koliaraki V, Pasparakis M & Kollias G (2015) IKKbeta in intestinal mesenchymal cells promotes initiation of colitis-associated cancer. *J Exp Med* **212**, 2235–2251.
- 72 Pallangyo CK, Ziegler PK & Greten FR (2015) IKKbeta acts as a tumor suppressor in cancer-associated fibroblasts during intestinal tumorigenesis. *J Exp Med* **212**, 2253–2266.
- 73 Fordham RP & Sansom OJ (2015) Colon contradictions: NF-kappaB signaling in intestinal tumorigenesis. *J Exp Med* **212**, 2185.
- 74 Guma M, Stepniak D, Shaked H, Spehlmann ME, Shenouda S, Cheroute H, Vicente-Suarez I, Eckmann L, Kagnoff MF & Karin M (2011) Constitutive intestinal NF-kappaB does not trigger destructive inflammation unless accompanied by MAPK activation. *J Exp Med* **208**, 1889–1900.
- 75 Lee SH, Hu LL, Gonzalez-Navajas J, Seo GS, Shen C, Brick J, Herdman S, Varki N, Corr M, Lee J *et al.* (2010) ERK activation drives intestinal tumorigenesis in Apc(min/+) mice. *Nat Med* **16**, 665–670.
- 76 Goktuna SI, Shostak K, Chau TL, Heukamp LC, Hennuy B, Duong HQ, Ladang A, Close P, Klevernic I, Olivier F *et al.* (2016) The prosurvival IKK-related kinase IKKepsilon integrates LPS and IL17A signaling cascades to promote Wnt-dependent tumor development in the intestine. *Cancer Res* **76**, 2587–2599.
- 77 Schwitalla S, Ziegler PK, Horst D, Becker V, Kerle I, Begus-Nahrmann Y, Lechel A, Rudolph KL, Langer R, Slotta-Huspenina J *et al.* (2013) Loss of p53 in enterocytes generates an inflammatory microenvironment enabling invasion and lymph node metastasis of carcinogen-induced colorectal tumors. *Cancer Cell* **23**, 93–106.
- 78 Feng Y, Santoriello C, Mione M, Hurlstone A & Martin P (2010) Live imaging of innate immune cell sensing of transformed cells in zebrafish larvae: parallels between tumor initiation and wound inflammation. *PLoS Biol* **8**, e1000562.
- 79 Burke F, East N, Upton C, Patel K & Balkwill FR (1997) Interferon gamma induces cell cycle arrest and apoptosis in a model of ovarian cancer: enhancement of effect by batimastat. *Eur J Cancer* **33**, 1114–1121.
- 80 Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ & Schreiber RD (2001) IFNgamma and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature* **410**, 1107–1111.
- 81 Munn DH & Cheung NK (1990) Phagocytosis of tumor cells by human monocytes cultured in recombinant macrophage colony-stimulating factor. *J Exp Med* **172**, 231–237.
- 82 Lawrence T, Bebien M, Liu GY, Nizet V & Karin M (2005) IKKalpha limits macrophage NF-kappaB activation and contributes to the resolution of inflammation. *Nature* **434**, 1138–1143.
- 83 Shembade N, Pujari R, Harhaj NS, Abbott DW & Harhaj EW (2011) The kinase IKKalpha inhibits activation of the transcription factor NF-kappaB by phosphorylating the regulatory molecule TAX1BP1. *Nat Immunol* **12**, 834–843.
- 84 Mancino A, Habbedine M, Johnson E, Luron L, Bebien M, Memet S, Fong C, Bajenoff M, Wu X, Karin M *et al.* (2013) I kappa B kinase alpha (IKKalpha) activity is required for functional maturation of dendritic cells and acquired immunity to infection. *EMBO J* **32**, 816–828.
- 85 Gloire G, Horion J, El Mjiyad N, Bex F, Chariot A, Dejardin E & Piette J (2007) Promoter-dependent effect of IKKalpha on NF-kappaB/p65 DNA binding. *J Biol Chem* **282**, 21308–21318.
- 86 Margalef P, Fernandez-Majada V, Villanueva A, Garcia-Carbonell R, Iglesias M, Lopez L, Martinez-

- Iniesta M, Villa-Freixa J, Mulero MC, Andreu M *et al.* (2012) A truncated form of IKK α is responsible for specific nuclear IKK activity in colorectal cancer. *Cell Rep* **2**, 840–854.
- 87 Hu MC, Lee DF, Xia W, Golfman LS, Ou-Yang F, Yang JY, Zou Y, Bao S, Hanada N, Saso H *et al.* (2004) IkappaB kinase promotes tumorigenesis through inhibition of forkhead FOXO3a. *Cell* **117**, 225–237.
- 88 Lee DF, Kuo HP, Chen CT, Hsu JM, Chou CK, Wei Y, Sun HL, Li LY, Ping B, Huang WC *et al.* (2007) IKK beta suppression of TSC1 links inflammation and tumor angiogenesis via the mTOR pathway. *Cell* **130**, 440–455.
- 89 Huber MA, Azoitei N, Baumann B, Grunert S, Sommer A, Pehamberger H, Kraut N, Beug H & Wirth T (2004) NF-kappaB is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression. *J Clin Invest* **114**, 569–581.
- 90 Wu Y, Deng J, Rychahou PG, Qiu S, Evers BM & Zhou BP (2009) Stabilization of snail by NF-kappaB is required for inflammation-induced cell migration and invasion. *Cancer Cell* **15**, 416–428.
- 91 Li CW, Xia W, Huo L, Lim SO, Wu Y, Hsu JL, Chao CH, Yamaguchi H, Yang NK, Ding Q *et al.* (2012) Epithelial-mesenchymal transition induced by TNF-alpha requires NF-kappaB-mediated transcriptional upregulation of Twist1. *Cancer Res* **72**, 1290–1300.
- 92 Magliozzi R, Low TY, Weijts BG, Cheng T, Spanjaard E, Mohammed S, van Veen A, Ovaas H, de Rooij J, Zwartkruis FJ *et al.* (2013) Control of epithelial cell migration and invasion by the IKKbeta- and CK1alpha-mediated degradation of RAPGEF2. *Dev Cell* **27**, 574–585.
- 93 Park BK, Zhang H, Zeng Q, Dai J, Keller ET, Giordano T, Gu K, Shah V, Pei L, Zarbo RJ *et al.* (2007) NF-kappaB in breast cancer cells promotes osteolytic bone metastasis by inducing osteoclastogenesis via GM-CSF. *Nat Med* **13**, 62–69.
- 94 Chen C, Cao F, Bai L, Liu Y, Xie J, Wang W, Si Q, Yang J, Chang A, Liu D *et al.* (2015) IKKbeta enforces a LIN28B/TCF7L2 positive feedback loop that promotes cancer cell stemness and metastasis. *Cancer Res* **75**, 1725–1735.
- 95 Yeh PY, Lu YS, Ou DL & Cheng AL (2011) IkappaB kinases increase Myc protein stability and enhance progression of breast cancer cells. *Mol Cancer* **10**, 53.
- 96 Romieu-Mourez R, Landesman-Bollag E, Seldin DC, Traish AM, Mercurio F & Sonenshein GE (2001) Roles of IKK kinases and protein kinase CK2 in activation of nuclear factor-kappaB in breast cancer. *Cancer Res* **61**, 3810–3818.
- 97 Park KJ, Krishnan V, O'Malley BW, Yamamoto Y & Gaynor RB (2005) Formation of an IKK α -dependent transcription complex is required for estrogen receptor-mediated gene activation. *Mol Cell* **18**, 71–82.
- 98 Dan HC, Antonia RJ & Baldwin AS (2016) PI3K/Akt promotes feedforward mTORC2 activation through IKK α . *Oncotarget* **7**, 21064–21075.
- 99 Bist P, Leow SC, Phua QH, Shu S, Zhuang Q, Loh WT, Nguyen TH, Zhou JB, Hooi SC & Lim LH (2011) Annexin-1 interacts with NEMO and RIP1 to constitutively activate IKK complex and NF-kappaB: implication in breast cancer metastasis. *Oncogene* **30**, 3174–3185.
- 100 Stylianou S, Clarke RB & Brennan K (2006) Aberrant activation of notch signaling in human breast cancer. *Cancer Res* **66**, 1517–1525.
- 101 Robinson DR, Kalyana-Sundaram S, Wu YM, Shankar S, Cao X, Ateeq B, Asangani IA, Iyer M, Maher CA, Grasso CS *et al.* (2011) Functionally recurrent rearrangements of the MAST kinase and Notch gene families in breast cancer. *Nat Med* **17**, 1646–1651.
- 102 Sethi N, Dai X, Winter CG & Kang Y (2011) Tumor-derived JAGGED1 promotes osteolytic bone metastasis of breast cancer by engaging notch signaling in bone cells. *Cancer Cell* **19**, 192–205.
- 103 Jin S, Mutvei AP, Chivukula IV, Andersson ER, Ramskold D, Sandberg R, Lee KL, Kronqvist P, Mamaeva V, Ostling P *et al.* (2013) Non-canonical Notch signaling activates IL-6/JAK/STAT signaling in breast tumor cells and is controlled by p53 and IKK α /IKK β . *Oncogene* **32**, 4892–4902.
- 104 Cao Y, Luo JL & Karin M (2007) IkappaB kinase alpha kinase activity is required for self-renewal of ErbB2/Her2-transformed mammary tumor-initiating cells. *Proc Natl Acad Sci USA* **104**, 15852–15857.
- 105 Bennett L, Quinn J, McCall P, Mallon EA, Horgan PG, McMillan DC, Paul A & Edwards J (2017) High IKK α expression is associated with reduced time to recurrence and cancer specific survival in oestrogen receptor (ER)-positive breast cancer. *Int J Cancer* **140**, 1633–1644.
- 106 Zhang W, Tan W, Wu X, Poustovoitov M, Strasner A, Li W, Borcherding N, Ghassemian M & Karin M (2013) A NIK-IKK α module expands ErbB2-induced tumor-initiating cells by stimulating nuclear export of p27/Kip1. *Cancer Cell* **23**, 647–659.
- 107 Merkhofer EC, Cogswell P & Baldwin AS (2010) Her2 activates NF-kappaB and induces invasion through the canonical pathway involving IKK α . *Oncogene* **29**, 1238–1248.
- 108 Tan W, Zhang W, Strasner A, Grivennikov S, Cheng JQ, Hoffman RM & Karin M (2011) Tumour-infiltrating regulatory T cells stimulate mammary cancer metastasis through RANKL-RANK signalling. *Nature* **470**, 548–553.

- 109 Mauad TH, van Nieuwkerk CM, Dingemans KP, Smit JJ, Schinkel AH, Notenboom RG, van den Bergh Weerman MA, Verkruijsen RP, Groen AK, Oude Elferink RP *et al.* (1994) Mice with homozygous disruption of the *mdr2* P-glycoprotein gene. A novel animal model for studies of nonsuppurative inflammatory cholangitis and hepatocarcinogenesis. *Am J Pathol* **145**, 1237–1245.
- 110 Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, Gutkovich-Pyest E, Urieli-Shoval S, Galun E & Ben-Neriah Y (2004) NF- κ B functions as a tumour promoter in inflammation-associated cancer. *Nature* **431**, 461–466.
- 111 Maeda S, Kamata H, Luo JL, Leffert H & Karin M (2005) IKK β couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell* **121**, 977–990.
- 112 Pham CG, Bubici C, Zazzeroni F, Papa S, Jones J, Alvarez K, Jayawardena S, De Smaele E, Cong R, Beaumont C *et al.* (2004) Ferritin heavy chain upregulation by NF- κ B inhibits TNF α -induced apoptosis by suppressing reactive oxygen species. *Cell* **119**, 529–542.
- 113 Sakurai T, Maeda S, Chang L & Karin M (2006) Loss of hepatic NF- κ B activity enhances chemical hepatocarcinogenesis through sustained c-Jun N-terminal kinase 1 activation. *Proc Natl Acad Sci USA* **103**, 10544–10551.
- 114 He G, Yu GY, Temkin V, Ogata H, Kuntzen C, Sakurai T, Sieghart W, Peck-Radosavljevic M, Leffert HL & Karin M (2010) Hepatocyte IKK β /NF- κ B inhibits tumor promotion and progression by preventing oxidative stress-driven STAT3 activation. *Cancer Cell* **17**, 286–297.
- 115 Kondylis V, Polykratis A, Ehlken H, Ochoa-Callejero L, Straub BK, Krishna-Subramanian S, Van TM, Curth HM, Heise N, Weih F *et al.* (2015) NEMO prevents steatohepatitis and hepatocellular carcinoma by inhibiting RIPK1 kinase activity-mediated hepatocyte apoptosis. *Cancer Cell* **28**, 582–598.
- 116 Naugler WE, Sakurai T, Kim S, Maeda S, Kim K, Elsharkawy AM & Karin M (2007) Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science* **317**, 121–124.
- 117 Liu M, Lee DF, Chen CT, Yen CJ, Li LY, Lee HJ, Chang CJ, Chang WC, Hsu JM, Kuo HP *et al.* (2012) IKK α activation of NOTCH links tumorigenesis via FOXA2 suppression. *Mol Cell* **45**, 171–184.
- 118 Haybaeck J, Zeller N, Wolf MJ, Weber A, Wagner U, Kurrer MO, Bremer J, Iezzi G, Graf R, Clavien P-A *et al.* (2009) A lymphotoxin-driven pathway to hepatocellular carcinoma. *Cancer Cell* **16**, 295–308.
- 119 Wolf MJ, Adili A, Piotrowitz K, Abdullah Z, Boege Y, Stemmer K, Ringelhan M, Simonavicius N, Egger M, Wohlleber D *et al.* (2014) Metabolic activation of intrahepatic CD8⁺ T cells and NKT cells causes nonalcoholic steatohepatitis and liver cancer via cross-talk with hepatocytes. *Cancer Cell* **26**, 549–564.
- 120 Endig J, Buitrago-Molina LE, Marhenke S, Reisinger F, Saborowski A, Schutt J, Limbourg F, Konecke C, Schreder A, Michael A *et al.* (2016) Dual role of the adaptive immune system in liver injury and hepatocellular carcinoma development. *Cancer Cell* **30**, 308–323.
- 121 Zhang K & Kaufman RJ (2008) From endoplasmic-reticulum stress to the inflammatory response. *Nature* **454**, 455–462.
- 122 Lee DF, Kuo HP, Chen CT, Wei Y, Chou CK, Hung JY, Yen CJ & Hung MC (2008) IKK β suppression of TSC1 function links the mTOR pathway with insulin resistance. *Int J Mol Med* **22**, 633–638.
- 123 Reid MA, Lowman XH, Pan M, Tran TQ, Warmoes MO, Ishak Gabra MB, Yang Y, Locasale JW & Kong M (2016) IKK β promotes metabolic adaptation to glutamine deprivation via phosphorylation and inhibition of PFKFB3. *Genes Dev* **30**, 1837–1851.
- 124 Cubillos-Ruiz JR, Bettigole SE & Glimcher LH (2017) Tumorigenic and immunosuppressive effects of endoplasmic reticulum stress in cancer. *Cell* **168**, 692–706.
- 125 Niederreiter L, Fritz TM, Adolph TE, Krismer AM, Offner FA, Tschurtschenthaler M, Flak MB, Hosomi S, Tomczak MF, Kaneider NC *et al.* (2013) ER stress transcription factor Xbp1 suppresses intestinal tumorigenesis and directs intestinal stem cells. *J Exp Med* **210**, 2041–2056.
- 126 Nakagawa H, Umemura A, Taniguchi K, Font-Burgada J, Dhar D, Ogata H, Zhong Z, Valasek MA, Seki E, Hidalgo J *et al.* (2014) ER stress cooperates with hypernutrition to trigger TNF-dependent spontaneous HCC development. *Cancer Cell* **26**, 331–343.
- 127 Li Q, Lu Q, Hwang JY, Buscher D, Lee KF, Izpisua-Belmonte JC & Verma IM (1999) IKK1-deficient mice exhibit abnormal development of skin and skeleton. *Genes Dev* **13**, 1322–1328.
- 128 Hu Y, Baud V, Delhase M, Zhang P, Deerinck T, Ellisman M, Johnson R & Karin M (1999) Abnormal morphogenesis but intact IKK activation in mice lacking the IKK α subunit of I κ B kinase. *Science* **284**, 316–320.
- 129 Takeda K, Takeuchi O, Tsujimura T, Itami S, Adachi O, Kawai T, Sanjo H, Yoshikawa K, Terada N & Akira S (1999) Limb and skin abnormalities in mice lacking IKK α . *Science* **284**, 313–316.
- 130 Hu Y, Baud V, Oga T, Kim KI, Yoshida K & Karin M (2001) IKK α controls formation of the epidermis independently of NF- κ B. *Nature* **410**, 710–714.

- 131 Sil AK, Maeda S, Sano Y, Roop DR & Karin M (2004) IkappaB kinase-alpha acts in the epidermis to control skeletal and craniofacial morphogenesis. *Nature* **428**, 660–664.
- 132 Liu B, Park E, Zhu F, Bustos T, Liu J, Shen J, Fischer SM & Hu Y (2006) A critical role for I kappaB kinase alpha in the development of human and mouse squamous cell carcinomas. *Proc Natl Acad Sci USA* **103**, 17202–17207.
- 133 Maeda G, Chiba T, Kawashiri S, Satoh T & Imai K (2007) Epigenetic inactivation of IkappaB Kinase-alpha in oral carcinomas and tumor progression. *Clin Cancer Res* **13**, 5041–5047.
- 134 Descargues P, Sil AK & Karin M (2008) IKKalpha, a critical regulator of epidermal differentiation and a suppressor of skin cancer. *EMBO J* **27**, 2639–2647.
- 135 Marinari B, Moretti F, Botti E, Giustizieri ML, Descargues P, Giunta A, Stolfi C, Ballaro C, Papoutsaki M, Alema S *et al.* (2008) The tumor suppressor activity of IKKalpha in stratified epithelia is exerted in part via the TGF-beta antiproliferative pathway. *Proc Natl Acad Sci USA* **105**, 17091–17096.
- 136 Kwak YT, Radaideh SM, Ding L, Li R, Frenkel E, Story MD, Girard L, Minna J & Verma UN (2011) Cells lacking IKKalpha show nuclear cyclin D1 overexpression and a neoplastic phenotype: role of IKKalpha as a tumor suppressor. *Mol Cancer Res* **9**, 341–349.
- 137 Liu B, Willette-Brown J, Liu S, Chen X, Fischer SM & Hu Y (2011) IKKalpha represses a network of inflammation and proliferation pathways and elevates c-Myc antagonists and differentiation in a dose-dependent manner in the skin. *Cell Death Differ* **18**, 1854–1864.
- 138 Park E, Zhu F, Liu B, Xia X, Shen J, Bustos T, Fischer SM & Hu Y (2007) Reduction in IkappaB kinase alpha expression promotes the development of skin papillomas and carcinomas. *Cancer Res* **67**, 9158–9168.
- 139 Liu B, Xia X, Zhu F, Park E, Carbajal S, Kiguchi K, DiGiovanni J, Fischer SM & Hu Y (2008) IKKalpha is required to maintain skin homeostasis and prevent skin cancer. *Cancer Cell* **14**, 212–225.
- 140 Xia X, Park E, Liu B, Willette-Brown J, Gong W, Wang J, Mitchell D, Fischer SM & Hu Y (2010) Reduction of IKKalpha expression promotes chronic ultraviolet B exposure-induced skin inflammation and carcinogenesis. *Am J Pathol* **176**, 2500–2508.
- 141 Xia X, Liu S, Xiao Z, Zhu F, Song NY, Zhou M, Liu B, Shen J, Nagashima K, Veenstra TD *et al.* (2013) An IKKalpha-nucleophosmin axis utilizes inflammatory signaling to promote genome integrity. *Cell Rep* **5**, 1243–1255.
- 142 Alameda JP, Moreno-Maldonado R, Fernandez-Acenero MJ, Navarro M, Page A, Jorcano JL, Bravo A, Ramirez A & Casanova ML (2011) Increased IKKalpha expression in the basal layer of the epidermis of transgenic mice enhances the malignant potential of skin tumors. *PLoS ONE* **6**, e21984.
- 143 Alameda JP, Navarro M, Ramirez A, Page A, Suarez-Cabrera C, Moreno-Maldonado R, Paramio JM, del Carmen Farina MR, Del Rio M, Fernandez-Acenero MJ *et al.* (2016) IKKalpha regulates the stratification and differentiation of the epidermis: implications for skin cancer development. *Oncotarget* **7**, 76779–76792.
- 144 Brown K, Buchmann A & Balmain A (1990) Carcinogen-induced mutations in the mouse c-Ha-ras gene provide evidence of multiple pathways for tumor progression. *Proc Natl Acad Sci USA* **87**, 538–542.
- 145 Wang XJ, Greenhalgh DA & Roop DR (2000) Transgenic coexpression of v-Ha-ras and transforming growth factor alpha increases epidermal hyperproliferation and tumorigenesis and predisposes to malignant conversion via endogenous c-Ha-ras activation. *Mol Carcinog* **27**, 200–209.
- 146 Malliri A, van der Kammen RA, Clark K, van der Valk M, Michiels F & Collard JG (2002) Mice deficient in the Rac activator Tiam1 are resistant to Ras-induced skin tumours. *Nature* **417**, 867–871.
- 147 Alameda JP, Gaspar M, Ramirez A, Navarro M, Page A, Suarez-Cabrera C, Fernandez MG, Merida JR, Paramio JM, Garcia-Fernandez RA *et al.* (2016) Deciphering the role of nuclear and cytoplasmic IKKalpha in skin cancer. *Oncotarget* **7**, 29531–29547.
- 148 Page A, Bravo A, Suarez-Cabrera C, Alameda JP, Casanova ML, Lorz C, Segrelles C, Segovia JC, Paramio JM, Navarro M *et al.* (2017) IKKbeta-mediated resistance to skin cancer development is Ink4a/Arf-dependent. *Mol Cancer Res* **15**, 1255–1264.
- 149 Ho YS, Chen CH, Wang YJ, Pestell RG, Albanese C, Chen RJ, Chang MC, Jeng JH, Lin SY, Liang YC *et al.* (2005) Tobacco-specific carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) induces cell proliferation in normal human bronchial epithelial cells through NFkappaB activation and cyclin D1 up-regulation. *Toxicol Appl Pharmacol* **205**, 133–148.
- 150 Takahashi H, Ogata H, Nishigaki R, Broide DH & Karin M (2010) Tobacco smoke promotes lung tumorigenesis by triggering IKKbeta- and JNK1-dependent inflammation. *Cancer Cell* **17**, 89–97.
- 151 Jackson EL, Willis N, Mercer K, Bronson RT, Crowley D, Montoya R, Jacks T & Tuveson DA (2001) Analysis of lung tumor initiation and progression using conditional expression of oncogenic K-ras. *Genes Dev* **15**, 3243–3248.
- 152 Dougan M, Li D, Neuberg D, Mihm M, Googe P, Wong KK & Dranoff G (2011) A dual role for the immune response in a mouse model of inflammation-associated lung cancer. *J Clin Invest* **121**, 2436–2446.

- 153 Xiao Z, Jiang Q, Willette-Brown J, Xi S, Zhu F, Burkett S, Back T, Song NY, Datla M, Sun Z *et al.* (2013) The pivotal role of IKK α in the development of spontaneous lung squamous cell carcinomas. *Cancer Cell* **23**, 527–540.
- 154 Ichikawa T, Suzuki H, Ueda T, Komiya A, Imamoto T & Kojima S (2005) Hormone treatment for prostate cancer: current issues and future directions. *Cancer Chemother Pharmacol* **56** (Suppl 1), 58–63.
- 155 Greenberg NM, DeMayo F, Finegold MJ, Medina D, Tilley WD, Aspinall JO, Cunha GR, Donjacour AA, Matusik RJ & Rosen JM (1995) Prostate cancer in a transgenic mouse. *Proc Natl Acad Sci USA* **92**, 3439–3443.
- 156 Luo JL, Tan W, Ricono JM, Korchynskiy O, Zhang M, Gonias SL, Cheresch DA & Karin M (2007) Nuclear cytokine-activated IKK α controls prostate cancer metastasis by repressing Maspin. *Nature* **446**, 690–694.
- 157 Ammirante M, Luo JL, Grivennikov S, Nedospasov S & Karin M (2010) B-cell-derived lymphotoxin promotes castration-resistant prostate cancer. *Nature* **464**, 302–305.
- 158 Ammirante M, Kuraishy AI, Shalapour S, Strasner A, Ramirez-Sanchez C, Zhang W, Shabaik A & Karin M (2013) An IKK α -E2F1-BMI1 cascade activated by infiltrating B cells controls prostate regeneration and tumor recurrence. *Genes Dev* **27**, 1435–1440.
- 159 Lesina M, Kurkowski MU, Ludes K, Rose-John S, Treiber M, Kloppel G, Yoshimura A, Reindl W, Sipos B, Akira S *et al.* (2011) Stat3/Socs3 activation by IL-6 transsignaling promotes progression of pancreatic intraepithelial neoplasia and development of pancreatic cancer. *Cancer Cell* **19**, 456–469.
- 160 Robinson-Smith TM, Isaacsohn I, Mercer CA, Zhou M, Van Rooijen N, Husseinzadeh N, McFarland-Mancini MM & Drew AF (2007) Macrophages mediate inflammation-enhanced metastasis of ovarian tumors in mice. *Cancer Res* **67**, 5708–5716.
- 161 Hernandez L, Hsu SC, Davidson B, Birrer MJ, Kohn EC & Annunziata CM (2010) Activation of NF- κ B signaling by inhibitor of NF- κ B kinase beta increases aggressiveness of ovarian cancer. *Cancer Res* **70**, 4005–4014.
- 162 Descargues P, Sil AK, Sano Y, Korchynskiy O, Han G, Owens P, Wang XJ & Karin M (2008) IKK α is a critical coregulator of a Smad4-independent TGF β -Smad2/3 signaling pathway that controls keratinocyte differentiation. *Proc Natl Acad Sci USA* **105**, 2487–2492.
- 163 Li N, Wu X, Holzer RG, Lee JH, Todoric J, Park EJ, Ogata H, Gukovskaya AS, Gukovsky I, Pizzo DP *et al.* (2013) Loss of acinar cell IKK α triggers spontaneous pancreatitis in mice. *J Clin Invest* **123**, 2231–2243.
- 164 Grivennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, Taniguchi K, Yu GY, Osterreicher CH, Hung KE *et al.* (2012) Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. *Nature* **491**, 254–258.
- 165 Baud V & Karin M (2009) Is NF- κ B a good target for cancer therapy? Hopes and pitfalls *Nat Rev Drug Discov* **8**, 33–40.
- 166 Tornatore L, Sandomenico A, Raimondo D, Low C, Rocci A, Tralau-Stewart C, Capece D, D'Andrea D, Bua M, Boyle E *et al.* (2014) Cancer-selective targeting of the NF- κ B survival pathway with GADD45 β /MKK7 inhibitors. *Cancer Cell* **26**, 495–508.
- 167 Greten FR, Arkan MC, Bollrath J, Hsu LC, Goode J, Miething C, Göktuna SI, Neuenhahn M, Fierer J, Paxian S *et al.* (2007) NF- κ B is a negative regulator of IL-1 β secretion as revealed by genetic and pharmacological inhibition of IKK β . *Cell* **130**, 918–931.
- 168 Eckmann L, Nebelsiek T, Fingerle AA, Dann SM, Mages J, Lang R, Robine S, Kagnoff MF, Schmid RM, Karin M *et al.* (2008) Opposing functions of IKK β during acute and chronic intestinal inflammation. *Proc Natl Acad Sci USA* **105**, 15058–15063.
- 169 Staudt LM (2010) Oncogenic activation of NF- κ B. *Cold Spring Harb Perspect Biol* **2**, a000109.
- 170 Nieto MA (2013) Epithelial plasticity: a common theme in embryonic and cancer cells. *Science* **342**, 1234850.
- 171 Dylla SJ, Beviglia L, Park IK, Chartier C, Raval J, Ngan L, Pickell K, Aguilar J, Lazetic S, Smith-Berdan S *et al.* (2008) Colorectal cancer stem cells are enriched in xenogeneic tumors following chemotherapy. *PLoS ONE* **3**, e2428.
- 172 Schreiber RD, Old LJ & Smyth MJ (2011) Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science* **331**, 1565–1570.
- 173 Zhang J, Yang PL & Gray NS (2009) Targeting cancer with small molecule kinase inhibitors. *Nat Rev Cancer* **9**, 28–39.
- 174 Garuti L, Roberti M & Bottegoni G (2010) Non-ATP competitive protein kinase inhibitors. *Curr Med Chem* **17**, 2804–2821.
- 175 Cao Y, Bonizzi G, Seagroves TN, Greten FR, Johnson R, Schmidt EV & Karin M (2001) IKK α provides an essential link between RANK signaling and cyclin D1 expression during mammary gland development. *Cell* **107**, 763–775.